

REMARKS

I. Status of the Claims

Claims 27, 28, 37-42 and 51-92 are pending in the application. Claims 37-42, 51-81, 83, 84 and 92 have been withdrawn from consideration by the Examiner as being drawn to a non-elected invention. No claims have been amended or added. By this amendment, no new matter has been added to the application.

II. Priority

Pursuant to the Examiner's request, the specification on page 1, in the paragraph beginning on line 6, has been amended to reflect that Ser. No. 10/214,286 is now issued U.S. Patent No. 6,852,737.

III. Claim Rejections Under 35 U.S.C. § 102(b)

Claims 27, 28, 82 and 85-91 are rejected as allegedly anticipated by Sartani *et al.*, U.S. Patent No. 5,767,136 ("Sartani") and Testa *et al.* (1997) *Cardiovascular Drug Reviews* 15(3):187-219 ("Testa"). The Examiner alleges that since both Sartani and Testa disclose lercanidipine, each reference anticipates the pending claims to Form II lercanidipine. The rejection is respectfully traversed, on the grounds that neither Sartani nor Testa discloses Form II lercanidipine.

Anticipation requires that every element set forth in a claim be disclosed explicitly or inherently in a single reference. The instant claims are directed to a crystalline polymorph of lercanidipine that is designated "Form II," and which is identifiable by its physical characteristics, e.g., characteristic peaks obtained upon X-ray diffraction. Neither Sartani nor Testa discloses a crystalline lercanidipine having X-ray diffraction peaks that are characteristic of Form II or having the other physical properties of the Form II polymorph. Thus, neither Sartani nor Testa explicitly discloses lercanidipine Form II lercanidipine. Neither does Testa or Sartani implicitly disclose lercanidipine Form II. Hence, neither Sartani nor Testa sets forth conditions for making lercanidipine that would be expected to yield Form II. Example 3 of Sartani cited by the Examiner includes only a general discussion of recrystallization of lercanidipine hydrochloride. Example 3, however, fails to provide any guidance as to which crystallization conditions would result in Form

polymorphs can be interconverted by heating or granulation. Id. at col. 10, lines 50-55.

U.S. Patent No. 5,412,095. Three polymorphs of terazosin monohydrochloride can be obtained from the same starting material (terazosin monohydrochloride methanolate) using the same solvent (ethanol). Terazosin Form I was obtained following dissolving terazosin monohydrochloride methanolate in hot absolute ethanol, cooling slowly to ambient temperature and standing overnight, and washing with dry acetone. (Ex. 5). Terazosin Form II was obtained by heating a slurry of terazosin monohydrochloride methanolate in absolute ethanol under reflux for approximately 24 h and cooling. (Ex. 6). Terazosin Form III was obtained by heating a slurry of terazosin monohydrochloride methanolate in absolute ethanol at 50°C for 30 min, followed by cooling in an ice bath and filtering. (Ex. 8).

▪ U.S. Patent No. 5,120,850. Describes obtaining different polymorphs of famotidine from the same solvents, depending on the cooling rate used during crystallization. Form A is obtained by starting with a hot solution and using a relatively slow cooling rate. '850 Patent at col. 2, lines 20-23. Form B is obtained by rapid cooling, which leads to rapid oversaturation. '850 Patent at col. 2, lines 23-29. Hence, Form A can be obtained by crystallization during slow cooling from boiling water or hot 50% methanol, 50% aqueous isopropanol, whereas Form B can be obtained from boiling water or hot 75% methanol, 50% aqueous isopropanol by placing the crystallization solution in an ice bath or pouring over ice (compare Ex. I/1, I/2 and I/4 to Ex. II/1, II/2, and II/3).

Thus, the foregoing examples illustrate that general guidance as to choice of solvents, cooling and drying conditions is not sufficient guidance to allow one of ordinary skill in the art to reproducibly obtain a particular polymorph. It cannot be presumed that all crystallization procedures falling within a general guidance for crystallization conditions will yield the same crystalline form.

With respect to the instant claims, the application makes clear that a simple reference to "crystalline lercanidipine" cannot be interpreted as a reference to Form II or indeed any other

particular crystalline form. The application discloses that it is possible to obtain at least four different crystalline forms of lercanidipine hydrochloride. Forms I and II are described in the application. Furthermore, the present specification discloses that lercanidipine hydrochloride crystalline Forms III and IV exist as well. *See* specification at page 13, lines 18-23. Hence, it is clear that crystalline lercanidipine hydrochloride can be present in several different physical forms. Each of these lercanidipine crystalline forms is obtainable by crystallization from, e.g., a “protic” solvent, depending on the precise conditions of crystallization and/or on the starting material.

With reference to the documents cited by the Examiner, Testa does not describe *any* crystallization conditions. Sartani does not describe crystallization conditions with such particularity that they would invariably lead to a particular lercanidipine crystalline form without the possibility of variation. Moreover, there will be sets of conditions that will have inherent variability. The Court of Appeals for the Federal Circuit has held that a prior art method for preparing crystalline forms does not anticipate a later-claimed crystalline form unless the method invariably leads to the claimed form. *Glaxo Inc. v Novopharm Ltd.*, 34 USPQ2d 1565, 52 F.3d 1043, 1047 (Fed. Cir. 1995), *cert. denied*, 516 US 988 (1995). There is no condition disclosed in Testa or Sartani that would lead invariably to Form II lercanidipine.

The present specification also dramatically illustrates that both lercanidipine hydrochloride Form I and lercanidipine hydrochloride Form II can be obtained following recrystallization from the same solvent, 2-propanol, depending on the conditions used. Example 4 of the present specification discloses preparation of Form I, by dissolving crude lercanidipine hydrochloride in 2-propanol under strong reflux and stirring, filtering, cooling to 40°C, maintaining the solution at 35°C for 24 h, then at 30°C for an additional 24 h, followed by filtering at 30°C, washing with 2-propanol, and drying at 70°C for 24 h. Example 10, by comparison, discloses preparation of Form II, by dissolving crude lercanidipine hydrochloride in a mixture of 2-propanol and water (8:2) at 60°C, filtering, cooling the solution to 25°C and stirring for 72 h at that temperature followed by collection of precipitate and drying. Thus, the instant specification itself illustrates the influence of crystallization conditions on the physical form that can be recovered. Both of the methods set forth in Examples 4 and 10 fall within the methods for “recrystallization of the crude [lercanidipine] hydrochloride compound from a solution of the compound in...a protic

solvent,” including isopropanol, and optionally including water that are set forth in Sartani (*see* column 7, lines 44-46, 56, and 61). Yet each method produces a different crystalline form.

Applicants submit that the foregoing is but one demonstration that methods falling within the teachings of Sartani can be used to obtain different polymorphs. Thus, Sartani cannot in any meaningful way disclose the “same” methods of preparation of crystalline lercanidipine hydrochloride that will invariably produce one form of lercanidipine or another. Furthermore, as discussed above, Testa does not disclose *any* method of preparing crystalline forms of lercanidipine. Accordingly, the claimed crystalline lercanidipine hydrochloride Form II is not necessarily described in Sartani or Testa, nor is it explicitly disclosed.

For the reasons set forth above, Applicants submit that claims 27, 28, 82 and 85-91 are not anticipated by Sartani or Testa. Reconsideration of these claims and withdrawal of the rejection thereof under 35 U.S.C. § 102(b) is requested.

IV. Claim Rejections Under 35 U.S.C. § 103(a)

Claims 27, 28, 82 and 85-91 are rejected as allegedly obvious over the combined teachings of Sartani and Testa in view of Haleblan *et al.* (1969) *J. Pharmaceutical Sci.* 58(8):911-929 (“Haleblan”); Chemical & Engineering News, Feb. 2003; Brittain *et al.* (1999) *Polymorphism in Pharmaceutical Sci.* pages 1-2, 185 (“Brittain”); Taday *et al.* (2003) *J. Pharm. Sci.* 92(4):831-838 (“Taday”); U.S. Pharmacopia #23, National Formulary #18 (1995); Muzaffar *et al.* (1979) *J. Pharmacy (Lahore)* 1(1):59-66 (“Muzaffar”); Jain *et al.* (1986) *Indian Drugs* 23(6):315-329 (“Jain”); and *Concise Encyclopedia Chemistry* (1993) 872-873.

The Examiner contends that Sartani and Testa teach crystal forms of lercanidipine, as well as pharmaceutical compositions. The Examiner further contends that Haleblan, Muzaffar, Jain, Taday and Brittain teach that compounds exist as polymorphs, and that Chemical & Engineering News, Muzaffar, the U.S. Pharmacopia and the Concise Encyclopedia of Chemistry teach that at any particular temperature and pressure, only one polymorph is stable. According to the Examiner, the claimed crystalline Form II and its properties are suggested by the cited references, and therefore, it would have been obvious to one of skill in the art in view of the references that lercanidipine would exist in different polymorphic forms.

incorrect storage or tablet preparation can affect the polymorphic state of a drug; and Doelker (2002) *Annales Pharmaceutiques Francaises* 60(3):161-176 as allegedly teaching that the environment of a polymorph can affect the polymorphic state. Furthermore, the Examiner cites Chemical & Engineering News as allegedly teaching that the formulation of drugs in metastable forms will cause the drug to convert to its most stable form and Otsuka *et al.* (1999) *Chem. Pharm. Bull.* 47(6):852-856 as allegedly teaching that a particular drug, carbamazepine, changes polymorphic form during preparation and formulation.

With regard to the enablement rejection, the Examiner contends that undue experimentation would be required to make pharmaceutical compositions containing crystalline Form II of lercanidipine. The Examiner bases her rejection on the content of the disclosure and the breadth of the claims, the level of unpredictability in the art, and the allegedly poor amount of direction provided in the specification.

Applicants respectfully traverse the rejection of claims 89-91 as allegedly lacking written description. Applicants submit that the Examiner has not met her burden of showing that the written description is inadequate. The description is presumed to be adequate unless or until sufficient evidence or reasoning to the contrary is presented to rebut the presumption. *See In re Marzocchi*, 439 F.2d 220, 224 (C.C.P.A. 1971). None of the references cited by the Examiner teaches that *lercanidipine Form II* changes to another polymorphic form during manufacture, formulation or storage. The cited references either provide a general teaching that a polymorphic form (and especially a metastable polymorphic form) of a chemical *may* change during manufacture, formulation or storage, or disclose examples of chemical compounds *other than* lercanidipine where a polymorphic form has changed.

As disclosed in the specification, lercanidipine Form II is a *stable* polymorph that has a high melting point (197-201°C), a lower solubility in aqueous media and in absolute ethanol compared to lercanidipine Form I, exhibits no weight loss up to its melting point in gravimetric analysis, and is non-hygroscopic. *See* specification at page 11, line 19-22; page 40, lines 15-17; page 42, lines 1-2; and Example 15, pages 42-43. It is well known in the art that crystalline solids generally make better active pharmaceutical ingredients ("API"). *See Remington: The Science and Practice of Pharmacy 20th ed.* (Alfonso R. Gennaro, ed., 2000), page 705 (attached hereto at Exhibit A). Furthermore, it is well known that stable polymorphs are usually desired for APIs because

metastable forms are prone to chemical and physical instability. *See, e.g.*, Exhibit A at page 706; Singhal & Curatolo (2003) *Adv. Drug Delivery Rev.* 56:335-347 at 336-337 (attached hereto at Exhibit B). Lercanidipine Form II is a stable polymorph that is desirable for pharmaceutical formulation. Furthermore, the specification discloses a number of suitable pharmaceutical excipients for use in the lercanidipine Form II pharmaceutical compositions. Therefore, the specification conveys with reasonable clarity to one skilled in the art that the applicants were in possession of the claimed pharmaceutical compositions.

For the reasons set forth above, Applicants submit that claims 89-91 are adequately described in the specification. Reconsideration of these claims and withdrawal of the rejection thereof under 35 U.S.C. § 112, first paragraph is requested.

Applicants also respectfully traverse the rejection of claims 89-91 as allegedly lacking enablement. First, as pointed out above with respect to the written description rejection, lercanidipine Form II is a *stable* polymorph, and a limited number of suitable pharmaceutical excipients for use in compositions containing Form II are disclosed in the specification. Second, pharmaceutical compositions containing a polymorphic form of an active ingredient and methods of making such compositions are well known in the art. *See, e.g., Physicians' Desk Reference* 58th ed. (Thomson 2004) for representative examples of drug formulations containing crystalline APIs in different formulations – TIAZAC[®], REMERONSolTab[®], ZITHROMAX[®], ZOLOFT[®] and AMBIEN[®] (attached hereto at Exhibit C). Moreover, a pharmaceutical composition (tablet) containing microcrystalline lercanidipine hydrochloride, lactose, microcrystalline cellulose, sodium starch glycolate, povidone and magnesium stearate is known and is available by prescription under the name ZANIDIP[®]. *See* ZANIDIP[®] prescribing information, available at <http://www.pbs.gov.au/pi/smpzanid31205.pdf>, last visited March 20, 2007 (attached hereto at Exhibit D). Thus, the Examiner has provided no reasonable basis to believe that the specification fails to enable the full scope of claims 89-91. The rejection should thus be withdrawn.

For the reasons set forth above, Applicants submit that claims 89-91 are enabled by the specification. Reconsideration of these claims and withdrawal of the rejection thereof under 35 U.S.C. § 112, first paragraph is requested.

VI. CONCLUSION

This application is believed to be in condition for allowance, which is earnestly solicited.

Dated: March 21, 2007

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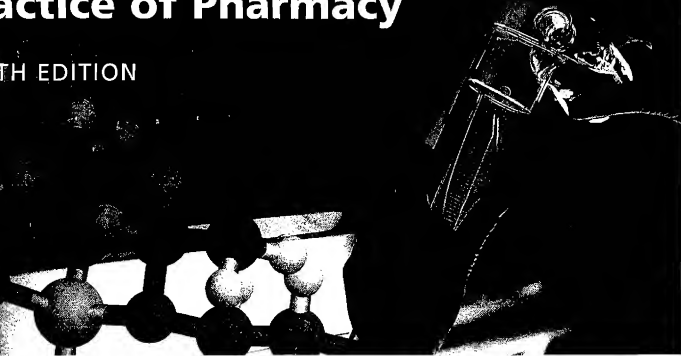
EXHIBIT A



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2 0 T H E D I T I O N

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Baltimore, Maryland 21201-2436 USA

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Philadelphia, PA 19106

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ISBN 0-683-306472

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2 3 4 5 6 7 8 9 10

pH-SOLUBILITY PROFILES

For a weak base, a plot of solubility versus pH will show the highest solubility at low pH and the lowest solubility at high pH; for weak acids, the opposite is true. Such plots give a graphic view of the impact of ionization on solubility for an NCE. The pH range of the small intestine, where oral absorption generally occurs, is approximately 6.5 to 8. It is undesirable to have a compound totally charged or uncharged in this region. If it is entirely charged, there are no un-ionized species that can be transported across the GI membrane. If it is totally uncharged, there are no charged species to enhance solubility. For a monoprotic NCE, the pK_a denotes the pH where the number of charged and uncharged species in solution are equal. On the ionized side of the pK_a , the solubility of the salt limits the maximum solubility. The solubility decline at very low pHs is due to activity and solubility-product effects.³⁻⁵ On the un-ionized side, the solubility of A^0 (the intrinsic solubility) marks the lowest solubility. Salts promote a saturated solution to be formed at a pH that is on the ionized side of the pK_a . They cannot alter the pK_a or the intrinsic solubility. Using these parameters, a qualitative pH-solubility profile can be constructed. Figure 38-5 shows pH-solubility profiles for different counter-acid salts.

The synthesis of salts depends on

1. A proton-exchange reactivity between A^0 and the counter-acid/base
2. A long-range order that permits crystal formation.

The discussion that follows will focus on forming salts from weak bases, because they comprise the majority of the new drug candidates. Weak acids would be treated analogously.

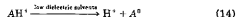
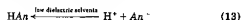
SALT-FORMING REACTIVITY POTENTIAL

In order for a salt to form, both the weak base, A^0 , and the counter-acid, HAn , must have sufficiently different pK_a values

such that a Brønsted-Lowry proton transfer from HAn to A^0 can take place. Table 38-2 gives potential counter-ions and their pK_a values from a listing of all drugs approved worldwide from 1983 to 1996. An acid-base proton transfer should be possible as long as the pK_a of HAn is less than that of the weak base A^0 (recall that the pK_a of A^0 is referenced to its protonated form A^0H^+ ; see *Solid-State Character*, page 702). If ΔpK_a is defined as

$$\Delta pK_a = pK_a(\text{weak base}) - pK_a(HAn) \quad (12)$$

a salt-forming reaction should be possible as long as ΔpK_a is positive. For example, a succinate salt (pK_a 4.2) with doxylamine (pK_a 4.4) is possible⁶ where the ΔpK_a is 0.2. Nevertheless, the greater the ΔpK_a , the greater the probability that a salt can be formed. Because the pK_a values in Table 38-2 are calculated for an aqueous environment, this rule must be used only as a guide for salt-forming reactivity in organic solvents. In an organic solvent in which the dielectric constant is lower than water, the ionization equilibria would be shifted:



For acridine bases, 50:50 ethanol:water weakens the aqueous pK_a by 1.41 pH units. For the counter-acid, HAn , pK_a weakening is greater than for the protonated base, A^0H^+ , because of the greater solubility of HAn in the organic phase and the production of two charges upon ionization. The net effect of organic solvent weakening is to reduce the pK_a difference between the counter-acid and the weak base. This lowers the salt-forming reactivity potential. Therefore, in a given organic solvent, if salt formation fails to occur for a particular aqueous ΔpK_a , it is unlikely that salts can be formed in this organic solvent with a smaller aqueous ΔpK_a .

VARYING SALT PROPERTIES USING COUNTER-ACID GROUPINGS

For weak bases, salt-forming counter-acids can be used to alter an API's solubility, dissolution, hygroscopicity, stability, and processing.⁶ Table 38-2 shows counter-acids organized into different functional groups. For each counter-acid, both the pK_a and the log P is given where appropriate. A starting point for salt expansion must begin with the properties of A^0 . If, for a weak base, $\Delta pK_a = pK_a^{A^0} - pK_{a, \text{counter-acid}, HAn} > 0$, then aqueous salts may be possible. Use of this table and the influence of different counter-acids are covered under *Decision-Tree, Goal-Oriented Approach*, page 712.

CRYSTAL FORMATION REQUIREMENTS

In general, crystalline solids, including salts, make the most promising APIs. The amorphous form of the solid state is usually not as stable as crystals, either physically or chemically. Crystal formation is a special characteristic of a solid in which the molecules self-organize into regular, repeating, molecular patterns. Solvents play at least three roles in crystallization.

1. They provide some solubilizing capacity so that concentrated solutions can be formed.
2. They promote the nucleation process. Nucleation may be from a pure solution (homogeneous nucleation) or from a seed crystal (heterogeneous nucleation). If a solvent binds too strongly to the molecular organizing functionalities of the salt or seed crystal, crystallization will be impeded. Finding appropriate solvents for crystal formation is a very important step in salt expansion. Failure to adequately explore and find solvents that can crystallize salts could mean that very usable salts would not be evaluated in the salt-selection step because they were not synthesized.

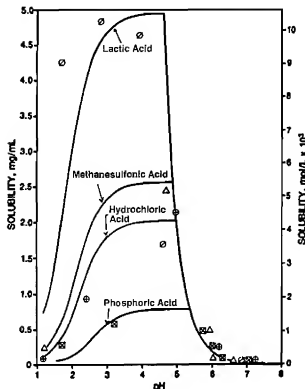


Figure 38-5. pH solubility profile of a weak base.³

3. Solvents, temperature, and cooling rate can impact the crystal-packing pattern of crystals. Stable polymorphic forms usually are desired for APIs. Metastable forms are normally avoided in an API because they are prone to physical and chemical instability. Solvent conditions that promote metastable and stable crystal formations will be explored under *Metastable Polymorph Formation*, page 710.

Salt Selection: Choosing the "Best" API

Salt selection is the first important API decision from the development perspective. Once a salt is chosen, time-consuming and lengthy toxicological studies are initiated that would have to be repeated if the salt form is changed. This decision involves choosing a solid-state phase, γA , which balances potentially conflicting needs: increasing absorption versus maintaining an API that is consistent and can be manufactured in a market-image dosage form (see *Compressibility and Compactibility*, page 712). Figure 38-6 shows some of the factors involved in this decision.

Permeability, solubility (C_S), and pK_a are intrinsic properties of A^0 that have been already determined in the analog selection phase (see Fig 38-4). The major dependent variables, absorption and consistency of the API, can be manipulated and balanced in salt selection. In the following sections, the impact of dissolution and particle size on absorption will be explored. In addition, the consistency of the API solid state under the influence of environmental destabilizing factors—such as exposure time (t), ultraviolet light (UV), pH, moisture (H_2O), temperature (T), and pharmaceutical processing operations like milling, compression, and compaction—will be considered.

ABSORPTION ASSESSMENT

Oral absorption is generally viewed as two-step, sequential process:

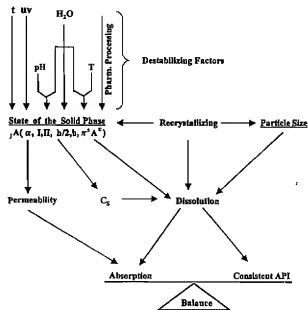
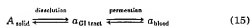


Figure 38-6. API salt selection decision: a balance between absorption and consistency.

Either dissolution of solid drug, A_{solid} , after the dosage form disintegrates in the GI tract, or the permeation of the dissolved drug, $a_{\text{GI tract}}$, through the GI membrane could be the slowest process. The slower of these two steps determines the overall rate of absorption and is thus rate-limiting.

Dissolution-limited absorption occurs when the rate of appearance in the GI tract by dissolution (a_{GI}) is slower than the rate of appearance in the systemic system (a_{blood}); *permeation-limited* absorption occurs when the a_{blood} appearance is the slowest process. The impact of these two rate processes on *in vitro-in vivo* (IVIV) correlations will be discussed in the section *Biopharmaceutical Classification of API*, page 714. Dissolution-limited absorption will now be considered.

The rate of dissolution of a particle is given by the Noyes-Whitney equation,

$$dA/dt = k_p S_p (C_S - C_{\text{bulk}}) \quad (\text{non-sink conditions}) \quad (16)$$

where

A is the amount of drug dissolved.

dA/dt is the rate of dissolution (Q sometimes is used for this rate).

k_p is the intrinsic dissolution constant for the drug.

S_p is the total surface area of the dissolving particle.

C_S is the saturation solubility of the drug at the surface of the particle.

C_{bulk} is the concentration of the drug in the bulk solution.

Because the rate of dissolution depends on the concentration difference between C_S and C_{bulk} , the maximum rate of dissolution would occur if $C_{\text{bulk}} = 0$ (ie, if drug was removed from solution as fast as it dissolved). This would be analogous to a sink that could drain the water coming out of a water faucet as fast as it comes in so that the water level never built up. This analogy is the basis for referring to Equation 16 as nonsink conditions for dissolution, because drug does build up in the solution and the rate of dissolution is correspondingly reduced.

The expression for the maximum dissolution rate is found by setting C_{bulk} equal to 0:

$$dA/dt = k_p S_p C_S \quad (\text{sink conditions}) \quad (17)$$

This initial rate of the Noyes-Whitney equation is termed sink conditions for the dissolution rate.

Particle-Size Effects—For a spherical drug particle of radius r , amount m , and of density ρ , Equation 17 can be rewritten as

$$dA/dt = (3k_p m/\rho) (1/r) C_S \quad (18)$$

This expression emphasizes the inverse relationship between the dissolution rate, dA/dt , and the particle size r , assuming no dissolution rate-reducing factors are present such as adsorbed air bubbles or aggregated particles.

Smaller particles dissolve faster than larger particles. Thus milling, a pharmaceutical unit-operation, increases dissolution because the API particle size is reduced. On the other hand, when drug particles are suspended in an aqueous solution, particles can increase in size due to recrystallization growth⁸ (Fig 38-7). Dosing such suspension orally would be expected to reduce absorption because of a reduction in the dissolution rate.

Reactive Media 1: Implications for Salts of Weak Acids and Weak Bases—When a drug reacts with gastric fluids, its dissolution deviates from Equation 17. For dissolution in 0.1 N HCl, acid-base reactivity is most important for salts of weak acids and for free bases. It has been found that the low pH environment of the stomach dissolves a salt of a weak acid 10 to 100 times faster than the weak acid itself.⁹ On the other hand, it is the free base, and not its HCl salt, that dissolves faster in this same environment.¹⁰ These deviations from Equation 17 have been shown to be due to differences between bulk-solution pHs and the pH at the surface of the drug particle. Thus, Equation 17 becomes

$$dA/dt = k_p S_p C_{SA} \quad (19)$$

EXHIBIT B

Drug polymorphism and dosage form design: a practical perspective

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Received 18 July 2003; accepted 6 October 2003

Abstract

Formulators are charged with the responsibility to formulate a product which is physically and chemically stable, manufacturable, and bioavailable. Most drugs exhibit structural polymorphism, and it is preferable to develop the most thermodynamically stable polymorph of the drug to assure reproducible bioavailability of the product over its shelf life under a variety of real-world storage conditions. There are occasional situations in which the development of a metastable crystalline or amorphous form is justified because a medical benefit is achieved. Such situations include those in which a faster dissolution rate or higher concentration are desired, in order to achieve rapid absorption and efficacy, or to achieve acceptable systemic exposure for a low-solubility drug. Another such situation is one in which the drug remains amorphous despite extensive efforts to crystallize it. If there is no particular medical benefit, there is less justification for accepting the risks of intentional development of a metastable crystalline or amorphous form. Whether or not there is medical benefit, the risks associated with development of a metastable form must be mitigated by laboratory work which provides assurance that (a) the largest possible form change will have no substantive effect on product quality or bioavailability, and/or (b) a change will not occur under all reasonable real-world storage conditions, and/or (c) analytical methodology and sampling procedures are in place which assure that a problem will be detected before dosage forms which have compromised quality or bioavailability can reach patients. © 2003 Elsevier B.V. All rights reserved.

Keywords: Amorphism; Dissolution; Polymorphism; Dosage form; Bioavailability; Stability; Mechanical properties

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1. Introduction

The subject of drug polymorphism has received extensive academic and industrial attention since the early pioneering reports of Aguiar and colleagues at Parke-Davis, in which effects of polymorphism on dissolution and bioavailability were highlighted for chloramphenicol palmitate [1,2]. Drug polymorphism has been the subject of hundreds of publications and numerous excellent reviews. For both an overview and an in-depth analysis of this complex field, see the excellent series of reviews in Volume 48 (2001) of *Advanced Drug Delivery Reviews* [4–9], in “Polymorphism in Pharmaceutical Sciences” edited by Brittain [10–19], and in “Solid State Chemistry of Drugs” by Byrn et al. [20]. In addition, two very clear reviews/commentaries from the regulatory perspective have appeared [21,22].

At this point in time, it would be difficult to say anything novel about the effects of polymorphism on physical stability, chemical stability, manufacturability, or oral absorption that has not been reviewed in the references quoted above. In many respects, the 1969 review by Halebian and McCrone was prescient in its broad coverage of the issues of polymorphism in pharmaceuticals [23]. In this article, we make no effort to review once again the vast literature on drug polymorphism. Furthermore, we do not here discuss theoretical or experimental details of the study of polymorphism. Rather, we attempt to provide a practical perspective on the impact of polymorphism on chemical stability, manufacturability, and bioavailability, with particular attention to a limited number of illustrative cases from our experience and the literature. Such a practical perspective must involve generalizations for which there are occasional exceptions.

2. Why develop multiple polymorphs?

It is generally accepted that, during the course of development of a drug, the lowest energy crystalline polymorph should be identified and chosen for development. This is critically important because the post-approval appearance of a polymorph with lower energy than the marketed polymorph can be catastrophic, as happened with the HIV protease inhibitor ritonavir [24]. For this reason, innovator pharmaceutical companies expend significant resources on this technical issue early in the development of a new drug. When executed carefully, the search for the lowest energy polymorph is arduous and time-consuming because (a) a variety of physical and chemical measurements must be made, and the stability of physical and chemical characteristics must be established in real-time storage models, (b) this search is not trivial, because a metastable polymorph may masquerade as the most stable form, and (c) every compound is different (i.e. the identity and properties of polymorphs are not theoretically predictable at present). The search for drug polymorphs is a complex empirical exercise, although recent advances in automation promise to make this activity somewhat less labor intensive.

There are three exceptions to the dictum that only the most stable polymorph should be developed. The first is extremely rare: the situation in which the lowest energy polymorph is chemically unstable due to the juxtaposition of two reactive groups in adjacent molecules in the crystal lattice. Such a “topochemical” reaction can in principle be avoided by identification of a crystalline polymorph in which the reactive species are no longer spatially close and/or oriented in a manner conducive to reaction. We are unaware of any examples of this phenomenon in

marketed drugs. The second exception is becoming more common, that is, the case of a drug whose absorption is solubility-limited and thus cannot achieve the systemic exposure required for therapy. In this case, a more soluble form of the drug is desired to deliver the therapeutic dose. The third exception is the situation in which it is desired to increase the dissolution rate of a drug to shorten T_{max} and/or increase C_{max} in order to bring quick relief for acute symptoms.

In the authors' opinion, when confronted with low solubility or the desire to decrease T_{max} or increase C_{max} , it is generally more productive to develop a stabilized amorphous form than a metastable crystalline polymorph. This will be discussed in more detail below.

In each of these three exceptions, a metastable polymorph or amorphous form is developed to provide a medical benefit.

If there is a desire to develop a metastable polymorph or amorphous form for a reason which does not provide a medical benefit, e.g. for manufacturing ease or for some other business reason, then the developer must assure that there is no significant risk to the patient. A rigorous laboratory-based analysis of the risks involved must be undertaken. This is of course also true when there is a medical benefit. In the sections below, we discuss the issues involved in the development of metastable polymorphs and amorphous forms, and their potential practical significance.

3. Chemical stability of polymorphs and amorphous forms

The polymorphs (or pseudopolymorphs) of some drugs have been shown to exhibit different chemical stability. Examples are carbamazepine [25], paroxetine maleate [26], indomethacin [27], methylprednisolone [28], furosemide [29], and enalapril maleate [30]. For example, the photodecay of form II of carbamazepine was 5- and 1.5-fold faster than forms I and III, respectively [25]. In addition to a change in the rate of decay, polymorphism may also affect the mechanism of decay, as observed in the reactivity of different polymorphs of cinnamic acid derivatives [31].

It is generally observed that the more thermodynamically stable polymorph is more chemically stable

than a metastable polymorph. This has generally been attributed to higher crystal packing density of the thermodynamically favored polymorph (i.e., the "density rule"), but recent investigation suggests that other factors, such as optimized orientation of molecules, and H-bonds and non-hydrogen bonds in the crystal lattice play a more important role. Relatively small changes in crystal packing may lead to significant differences in the crystal packing density and chemical reactivity of two polymorphs, as indomethacin polymorphs [27]. Indomethacin can exist as the metastable α -form and thermodynamically favored γ -form. As an exception to the density rule, the density of metastable α -form (1.42 g/mL) is higher than that of the γ -form (1.37 g/mL), suggesting tighter packing of the less stable polymorph. Although the metastable α -form has higher density, the α -form rapidly reacts with ammonia vapor while the γ -form is inert to ammonia. The lack in correlation between higher packing density and lower reactivity of the indomethacin polymorphs is due to the differences in crystal packing/hydrogen bonding. Higher density of the α -form is due to the presence of one extra H-bond in the crystal lattice. The differences in H-bonding and the crystal packing (two centrosymmetric carboxylic groups in α -form vs. three asymmetric molecules in γ -form) leads to a layer motif in the α -form that exposes the reactive carboxylic acid group to the crystal face, while in the γ -form, H-bonded carboxylic acid groups are buried in a hydrophobic cage. Easy accessibility of the reactive carboxylic acid groups in the α -form combined with the weak H-bond of one carboxylic acid group leads to higher reactivity of the α -form [27].

The intrinsic difference in chemical stability between two polymorphs, e.g. α - and γ -indomethacin, cannot be overcome, but a less chemically stable polymorph can often be formulated in a way which results in acceptable shelf-life.

In comparison to crystalline polymorphs, the amorphous form of a drug is generally expected to be less chemically stable due to the lack of a three dimensional crystalline lattice, higher free volume and greater molecular mobility. The chemical stability of amorphous systems has been discussed in detail elsewhere [20,32–35]. As early as 1965, amorphous penicillin G was shown to be less stable than the crystalline sodium and potassium salts [36]. Physical

change of amorphous molecules from a glassy state (at $T < T_g$) to a more mobile supercooled liquid state (at $T > T_g$) may further decrease chemical stability. For example, Asn-hexapeptide was found to be 10–100-fold more stable in the glassy state compared to its supercooled liquid state [37,38]. In addition to higher reactivity, the mechanism of degradation may be different in crystalline versus disordered materials. For example, methyl transfer was the major reaction pathway in unmilled crystalline tetraglycine methyl ester (TGME), while polycondensation was the major reaction pathway in milled TGME [39]. This change in mechanism from methyl transfer to polycondensation upon milling may be due to the creation of a disordered state with higher free volume where molecules can undergo the much higher change in orientation that is needed for the polycondensation reaction [39].

It should be pointed out that a major portion of any formulation effort is the choice of excipients and processes which minimize the chemical instability of the drug. If a metastable polymorph (or amorphous form) is less chemically stable than the lowest energy form of the drug, then in many cases it will be possible to maximize the chemical stability of this metastable form through judicious formulation decisions [40–45]. Thus reduced chemical stability of a metastable crystalline or amorphous drug form does not necessarily preclude its development as a product.

For a more in-depth review of chemical stability and drug physical state, see Bym et al. [9,20].

4. Mechanical properties of polymorphs and amorphous drug forms

Polymorphism can affect the mechanical properties of drug particles, and thus may impact the manufacturability and physical attributes of tablets. For example, polymorphs of metoprolol tartrate [46], paracetamol [47–50], sulfamerazine [51], phenobarbitalone [52], carbamazepine [53,54], phenylbutazone [55] and other drugs have been shown to exhibit different mechanical properties. A common effect of polymorphism is alteration of powder flow due to the difference in particle morphology of two polymorphs. Polymorphs with needle- or rod-shaped particles may have poor flow compared to polymorphs with low

aspect ratio, e.g. cubic habit or irregular spheres. The effect of polymorphism on other mechanical properties, such as hardness, yield pressure, elasticity, compressibility and bonding strength is more complex.

A simple general rule, although semi-empirical, proposed more than 20 years ago by Summers et al. can be used to predict the effect of crystal packing of polymorphs on their compressibility and bonding strength [55,56]. The more stable polymorph, due to its higher packing density, is expected to form stronger interparticle bonds but is harder to deform [46,55,56]. Since an increase in the bonding surface area resulting from deformation of particles may have higher impact on tablet strength than interparticle bond strength, the more stable of two polymorphs may provide weaker tablets. The mechanical properties of two enantiotropic polymorphs of metoprolol tartrate, metastable form I and the more stable form II (at room temperature), are consistent with this rule [46]. The porosity of pure drug tablets and yield pressure for form I were lower than for form II, suggesting that the less dense metastable form I may have less strength in the crystal lattice and be easier to deform. Form I also had higher elastic recovery, probably due to higher elasticity of form I and/or lower porosity of the tablets. As predicted, the tablets of the metastable form I were stronger at low pressures than those of form II, probably due to the higher compressibility of form I.

Factors other than those accounted for by the general rule proposed by Summers et al. may also affect the mechanical properties of two polymorphs. For example, the presence of slip planes in form I of sulfamerazine was found to be the reason for its higher plasticity than form II, the more stable form at room temperature [51]. This higher plasticity results in greater compressibility and tabletability. The authors of this study generalized this observation and suggested that crystals with slip planes would be expected to have superior tableting performance [51]. Recently, a fundamental atom–atom potential model simulation was used to predict a few mechanical properties of sulfathiazole and carbamazepine polymorphs [53]. More fundamental research in this area will improve our ability to predict the effect of polymorphism on mechanical properties.

For amorphous drug forms, mechanical properties may be different from those of crystalline drug due to

the absence of long range packing. The mechanical attributes of amorphous forms are less well understood than those of crystalline polymorphs. The lack of information on mechanical properties of amorphous drugs may be due to the physical and chemical instability of these forms, leading to reluctance in developing an amorphous form for a commercial drug product. Thus, an evaluation of mechanical properties of amorphous drugs is not routinely investigated in the pharmaceutical industry. One report comparing the mechanical properties of crystalline and amorphous forms of a model drug was published last year [57]. Compacts of amorphous material had higher brittleness and elasticity, and lower ductility than compacts prepared with the crystalline form.

Differences in the mechanical properties of two polymorphs or amorphous versus crystalline forms may or may not affect the manufacturability and physical attributes of tablets. For example, in the case of metoprolol tartrate, the differences in the mechanical properties of two polymorphs did not affect the bonding properties of tablets with relatively high drug loading [46]. The extent of the difference in the mechanical properties of two polymorphs, the drug loading, the robustness of each manufacturing step and the absolute value of the mechanical property undergoing change may be important parameters to consider while assessing the impact of polymorphism on manufacturability and physical attributes of tablets.

In some cases the favorable mechanical properties of one polymorph, even a metastable one, may be used to develop a more desirable process to manufacture tablets. For example, direct compression may be used to manufacture tablets with the more compressible orthorhombic form II of paracetamol instead of using more resource intensive granulation processes for monoclinic form I [47,50]. However, development of a metastable form for processing advantage should only be undertaken for drugs for which a very complete understanding exists with respect to form-dependent chemical stability, physical stability, and most importantly, bioavailability. This will typically be the case only for very old, highly studied, drugs.

As discussed above for chemical stability, manufacturability deficits of a particular polymorph may be overcome through judicious selection of excipients and processes. If a stable polymorph has problematic

mechanical properties, this certainly does not preclude its development. It is much more preferable to use excipients and processing to overcome the mechanical deficits of a stable polymorph than to develop an unstable polymorph because of its better mechanical properties.

For a review of the effects of processing (e.g. tableting) on drug form, see Morris et al. [8] and Brittain and Fiese [17]. For a discussion of the use of excipients to compensate for the physical properties of drugs in formulations, see Amidon [58].

5. Bioavailability of polymorphs

There are many reports of polymorph-dependent bioavailability and/or absorption rate, with much of this work done in animals. See for example animal studies of chloramphenicol palmitate [59], phenylbutazone [60], amobarbital [61], cimetidine [62], 6-mercaptopurine [63], and chlortetracycline [64]. For the purpose of the present analysis, we consider only human studies in detail.

5.1. Effects of polymorphism on dissolution and oral drug absorption in humans

Among the best known cases involving human dosing are those of chloramphenicol palmitate, mefenamic acid, oxytetracycline, and carbamazepine. These observations are quite old, having been reported in the 1950s and 1960s. For example, Aguiar et al. [1] demonstrated that absorption of chloramphenicol palmitate polymorph B was significantly greater than absorption of polymorph A in humans. Peak chloramphenicol serum levels were linearly proportional to the percentage of Form B in Form A/Form B mixtures. Chloramphenicol palmitate is a prodrug of chloramphenicol, which was prepared to provide a tasteless derivative [65]. Glazko et al. [66] reported that chloramphenicol palmitate must be hydrolyzed by intestinal esterases before the drug could be absorbed. Aguiar and colleagues demonstrated that *in vitro* hydrolysis of this prodrug by pancreatin was polymorph dependent, with significant hydrolysis of polymorph B and little hydrolysis of polymorph A. Aguiar and Zelmer [2] demonstrated that Form B dissolves faster than Form A, and has a much higher

solubility. This solubility difference probably results in the difference in ester hydrolysis rates, and ultimately the difference in oral absorption.

Aguiar and Zelmer [2] also reported on human absorption of two polymorphs of mefenamic acid. In this case, the two polymorphs gave almost identical blood levels. Aguiar and Zelmer calculated a free energy difference (ΔG_T) of -251 cal/mol between the two mefenamic acid polymorphs, where

$$\Delta G_T = RT \ln (\text{Solubility A/Solubility B})$$

In a similar manner, they calculated a free energy difference of -774 cal/mol between polymorphs A and B of chloramphenicol palmitate. These authors pointed out the correlation between the free energy difference and the observation of a polymorph-derived bioavailability difference (seen for chloramphenicol palmitate but not for mefenamic acid). However, the situation is clearly complicated by the issue of hydrolysis of the palmitate moiety in the lumen for chloramphenicol palmitate.

Brice and Hammer [67] reported in 1969 that oral dosing of 16 lots of oxytetracycline capsules from 13 suppliers gave drug blood levels which were lower than the innovator product. Seven of the lots gave oxytetracycline blood levels which were lower than the generally accepted minimum therapeutic level. Blood levels were generally correlated with in vitro dissolution rate. Groves subsequently reported large differences in in vitro dissolution performance of oxytetracycline tablets from various sources [68]. These studies made no attempt to relate dissolution observations to oxytetracycline polymorphism, and the observed differences may have resulted from differing formulations rather than differing polymorphs. Recently, Liebenberg et al. [69] compared six bulk oxytetracycline samples which met USP specifications, and noted that four of these contained one polymorph while the other two contained a different polymorph (form A). Tablets prepared from the form A polymorph dissolved significantly more slowly than the others in 0.1 M HCl. For example, the form A tablets exhibited $\sim 55\%$ dissolution at 30 min, while the others exhibited complete ($\sim 95\%$) dissolution at 30 min.

The drug carbamazepine exhibits polymorphism and product-to-product dissolution and bioavailability

ty differences, but a connection between these phenomena has not been directly experimentally demonstrated. Kahela et al. [70] reported that the anhydrous and dihydrate forms of carbamazepine exhibited very similar pharmacokinetics in humans. While the anhydrous form exhibited slower in vitro dissolution than the dihydrate in 0.1 M HCl, inclusion of 0.01% polysorbate 80 in the dissolution medium essentially eliminated this difference. Another study by Jumao-as et al. [71] demonstrated no difference in bioavailability between a generic carbamazepine product and the innovator product. Regardless, carbamazepine therapy with some products has been reported to be problematic [72,73]. Meyer et al. [74] reported on in vitro/in vivo studies of three out of 53 batches of generic carbamazepine tablets which were recalled due to clinical failures and dissolution changes. In vitro dissolution testing, carried out in water containing 1% sodium lauryl sulfate, revealed that two of the batches dissolved more slowly than the innovator product, and one batch dissolved more quickly. While the innovator product gave $\sim 95\%$ dissolution in 90 min in this medium, the slower generic batches gave $\sim 35\%$ and 75% dissolution. In humans, the generic batches gave mean relative AUCs (relative to the innovator) of 60–113%, with the same rank order observed in the in vitro dissolution behavior. It was suggested that moisture uptake during storage and particle size differences may have been involved in the irreproducible behavior of the generic tablets of this practically insoluble drug. It is known that anhydrous carbamazepine converts to the dihydrate quickly, e.g. completely within 1 h, when the anhydrous form is suspended in water [75].

The mechanistic uncertainty in these examples (i.e. whether drug physical form was involved in the observed dissolution or bioavailability differences) results from the lack of spectroscopic data which can identify the drug polymorph in a complex dosage form. Modern techniques such as ss-NMR and NIR can identify polymorphs in dosage forms (within limits), and should facilitate increased mechanistic understanding in future studies.

It is clear that for some drugs, there will be polymorph-dependent bioavailability. For a larger group, there will be polymorph-dependent absorption rate, reflected in in vivo C_{max} . For some pairs of polymorphs, there will be pharmacokinetic bioequi-

valence. As described above, in 1969 Aguiar and Zolmer proposed that polymorphs with a large free energy difference between them are likely to differ in pharmacokinetic behavior. This simply reflects a difference in solubility. In addition, polymorphs may exhibit different dissolution rates because of their different crystal habits, and this may also contribute to in vivo absorption rate differences.

For an excellent in-depth review of the relationships between polymorphism and solubility and dissolution rate, see Brittain and Grant [16].

5.2. The role of dose in bioavailability of high energy polymorphs

A significant solubility difference between two polymorphs is likely to result in a difference in oral absorption rate, reflected in a difference in C_{\max} . Differences in AUC, or oral bioavailability, will occur less often, and will depend upon the same underlying principles which govern the bioavailability differences between two unrelated drugs. Drug absorption may be modeled in a variety of ways [76,77]. A simple context in which to discuss this issue is provided by the concept of the maximum absorbable dose (MAD) [3,78]. The MAD is a conceptual tool which represents the quantity of drug which could be absorbed if the small intestine could be saturated with drug for 4.5 h (270 min), the average small intestinal transit time.

$$\text{MAD} = S \times K_a \times \text{SIWV} \times \text{SITT}$$

S , solubility (mg/ml) at pH 6.5; K_a , transintestinal absorption rate constant (min^{-1}); SIWV, small intestinal water volume (ml); SITT, small intestinal transit time (min).

The solubility at pH 6.5 reflects the solubility in the small intestine. K_a is determined in a rat intestinal perfusion experiment. In our laboratories, it has been observed that the human K_a is 1.4 times the rat K_a [79]. SIWV is the amount of water available for dissolution, generally accepted to be ~250 ml. While SIWV and SITT are approximations, moderate variations in these parameters do not significantly affect this analysis. The resulting MAD is in mg. This analysis ignores first pass intestinal and hepatic metabolism, which can be saturated, thus affecting bioavailability.

Table 1
MAD

Rat K_a (min^{-1}) (Human K_a)	Solubility (mg/ml)	MAD (mg) (Human)
0.003 [0.004]	0.01	2.7
0.003 [0.004]	0.02	5.4
0.003 [0.004]	0.03	8.1
0.03 [0.04]	0.01	27
0.03 [0.04]	0.02	54
0.03 [0.04]	0.03	81

If the intent is to increase bioavailability, it can be readily seen that increasing drug solubility will result in increased MAD (Table 1). In general, the range of solubility differences between polymorphs is typically 2–3-fold, due to the relatively small difference in free energy between polymorphs. Thus a higher energy polymorph with a solubility which is $3 \times$ that of the lowest energy polymorph may give a systemic exposure which is $3 \times$ that given by the low energy polymorph. As shown in Table 1, for a low human K_a of 0.004 min^{-1} , and a solubility of 0.01 mg/ml, a 3-fold increase in solubility only results in a MAD of 8.1 mg, which would be inadequate if the desired absorbed dose were, say, 50 mg. If bioavailability were practically governed in this way, there would not be much opportunity to increase the bioavailability of low-solubility drugs by developing a high energy polymorph or amorphous form.

In fact, equilibrium solubility may not be very relevant for oral absorption enhancement if polymorphs (or pseudopolymorphs) are physically unstable in the aqueous environment. Instead, intrinsic dissolution rate (IDR) and kinetic solubility over 4–6 h may be more relevant parameters to consider while studying the oral absorption of polymorphs. Form changes may sometimes occur during IDR and kinetic solubility measurements, but these changes are occurring on a timescale relevant for oral absorption, i.e. the small intestinal transit time. The kinetic solubility of a metastable polymorph over 4–6 h is often higher than its equilibrium solubility. The rank order of the IDR of polymorphs has been found to correlate well with the rank order for oral absorption due to the faster rate of dissolution of the less stable polymorph, leading to higher concentration of drug in solution available for absorption. Generally, this may

lead to a higher *in vivo* C_{\max} , but not a higher AUC, unless the drug is present in suspension throughout its small intestinal transit time (i.e. the dose is substantially greater than the MAD calculated for the thermodynamically stable polymorph). In some circumstances, the IDR and the achievable metastable supersaturation may temporarily provide a maximum drug concentration in the intestinal lumen which is in excess of the equilibrium solubility of the high energy polymorph. If the drug does not rapidly precipitate in the GI lumen, then the achievable MAD can conceivably be very large.

Although IDR may be a good single parameter to describe relative dissolution rates of two polymorphs, this does not take into account other factors that may govern oral absorption, namely, rate of conversion of one polymorph to another less soluble polymorph in the GI lumen, and the resulting precipitation of drug in the GI fluid. It is generally not possible to theoretically predict the degree of supersaturation of drug from a metastable polymorph or amorphous form, or the kinetics of physical conversion of one polymorph to another. However, these processes may be quantified by comparing the extent of supersaturation in model GI fluid (2) according to Eq. (1), and more importantly Eq. (2):

Supersaturated concentration ratio (SCR)

$$= C_{\max, \text{form 1}} / C_{\max, \text{form 2}} \quad (1)$$

Supersaturated AUC ratio (SAR)

$$= \text{AUC}_{\text{form 1}} / \text{AUC}_{\text{form 2}} \quad (2)$$

where $C_{\max, \text{form 1}}$ and $C_{\max, \text{form 2}}$ are the *in vitro* maximum concentrations of drug in solution from forms 1 and 2, respectively; and $\text{AUC}_{\text{form 1}}$ and $\text{AUC}_{\text{form 2}}$ are the areas under the *in vitro* drug concentration versus time curve over, say, 6 h, for forms 1 and 2, respectively. If a high dose (in substantial excess of the MAD for the stable polymorph) is dosed, and supersaturation is maintained for a long time, e.g. 6 h, while drug is absorbed, then the potential exists to achieve absorption of an amount of drug much higher than the MAD for the stable polymorph.

The greatest effect of dissolution rate and supersaturation of drug from a polymorph or amorphous

form is expected for compounds with high permeability and low solubility relative to dose (i.e. BCS class II compounds, where the administered dose will remain as a suspension for most of the absorption period). For solutes where the dose is expected to be very soluble in the GI fluid (i.e. BCS class I and III compounds) there may be no, or minimal differences in the AUC of polymorphs because solubility is not expected to be rate limiting in oral absorption.

5.3. Potential effects of physical instability of a metastable polymorph on oral absorption

Developing a bioequivalent product with a metastable form may not be easy, but in some cases it may be possible using formulation methods to achieve a bioequivalent AUC. It may be trickier, but possible, to blunt the higher pharmacokinetic C_{\max} which results from the higher dissolution rate of the metastable form. Thus, it may be possible to develop a formulation with a metastable drug form which is bioequivalent to the innovator formulation containing the thermodynamically most stable form. For some drugs, there is a potential danger that bioavailability could be lost if the metastable form converts to the more stable form during the shelf-life of the product. This is illustrated in Table 2. A metastable drug form may be formulated in a product (e.g. tablet) which has the same dissolution rate (Y) as a formulation of the stable drug form. Of course the metastable drug product will have to be formulated in a way which slows the drug dissolution rate. If the metastable form converts to the stable form in the product on storage, then the dissolution rate may decrease and *in vivo* performance may be compromised. This compromised *in vivo* performance may involve increased pharmacokinetic

Table 2
Potential performance changes on storage of a dosage form containing a metastable drug form

Drug form in formulation	IDR	Dissolution rate in formulated product	Dissolution rate in product after storage if metastable form converts to stable form
Metastable	$X + \Delta X$	Y	$Y - \Delta Y$
Stable	X	Y	Y

variability and, more extremely, decreased C_{max} and bioavailability.

As an example, phenylbutazone Form C exhibits a dissolution rate and solubility which are $1.5 \times$ and $1.2 \times$ that of Form A, respectively [80]. On storage at 40°C for 12 months, Form C was converted to 60% Form A. As another example, various marketed tablet formulations of glibenclamide have been shown to exhibit differing *in vitro* dissolution [81]. Glibenclamide exhibits forms which differ greater than 10-fold in solubility in simulated gastric fluid [82]. However, for glibenclamide the connection between product-to-product variability and polymorphism has not been directly demonstrated, but provides a possible explanation.

6. Dosage form decision

6.1. Metastable crystalline polymorph versus amorphous form

As discussed above, metastable crystalline polymorphs and amorphous forms may be less chemically stable and potentially possess different (in some cases less desirable) mechanical properties than the related stable crystalline form. These potential problems can in theory be solved by judicious choice of excipients and appropriate formulation strategies. In addition to chemical instability and mechanical properties, physical stability of the drug during product shelf life is of paramount importance in developing a drug product. A change in physical form can not only affect chemical stability and mechanical attributes of tablets, but much more importantly can compromise the oral absorption of a drug via a change in solubility.

Physical stabilization of intrinsically physically unstable crystalline polymorphs is a challenge because, by definition, the use of additives for improvement of physical stability involves a two phase system (polymorph and stabilizer) where the drug molecules are not in intimate contact with the stabilizer. Furthermore, physical conversion can be relatively precipitous, and exceptional care must be taken to design stability studies which cover all reasonable real-world conditions which such a formulation may encounter (e.g. temperature cycling). There is a need for increased understanding of stabilization of metastable

crystalline forms, and research in this area is sorely needed if practical solutions are to be found.

Amorphous forms are of course also physically unstable. For an introduction to the literature and general concepts on the physical stability of amorphous forms see Yu [5], Yoshioka et al. [83], and Crowley and Zografi [84]. Physical stabilization of amorphous forms is possible in some situations by generating intimate contact between the amorphous drug and the stabilizer by creating a drug/stabilizer dispersion [85–88]. The use of such dispersions, particularly with polymers, to intentionally enhance drug solubility has been known for many years [89,90], and practical formulations which achieve facile low-solubility drug dissolution and supersaturation have recently been described [91,92]. The identification of pharmaceutically acceptable stabilizers and processes which can inhibit solid state crystallization for a reasonable shelf-life is also a recent development [86].

While stabilized amorphous forms can sometimes be developed for intentional bioavailability improvement, the use of such forms to provide a dosage form which is bioequivalent to the stable drug crystalline form would be difficult, but perhaps possible in certain situations.

7. Solvates and hydrates

In general, the analysis provided above for the behavior of polymorphs also applies to metastable solvates and hydrates. For example, the dissolution rate and solubility of a drug can differ significantly for different solvates. Glibenclamide has been isolated as pentanol and toluene solvates, and these solvates exhibit higher solubility and dissolution rate than two non-solvated polymorphs [93]. In formulation of solvates (other than hydrates), the formulator must be careful to address the toxicity of the associated solvent, and carefully evaluate interactions of the drug and mobile solvent molecules with excipients on storage, which may result in compromised performance.

Similar to polymorphs in general, the physical stability of hydrates and anhydrous forms may depend upon the relative humidity and/or temperature of the environment, and the most stable form may switch as

the humidity/temperature is varied. Anhydrous to hydrate transitions can occur during dissolution at the drug/medium interface and can affect dissolution rate and perhaps bioavailability. Discussion of these issues is beyond the intended scope of this review.

Pharmaceutical solvates and hydrates have been reviewed by Morris [13], and hydrates have been reviewed by Khankari and Grant [94].

8. Conclusions

In principle, any polymorph or hydrate/solvate or amorphous form of a drug can be appropriately formulated. In practice, for some drugs constraints may be encountered. In general, the following conclusions are drawn from the literature and the experience of the authors:

1. It is always advisable to identify the lowest energy crystalline polymorph of a drug candidate during development, and to develop this form. While this form may not be the most processable form available, processing deficits can almost always be overcome with judicious choice of excipients and formulation processes. The lowest energy polymorph is almost always the most chemically stable form, and will not convert to another polymorph during storage as drug product. Of course, care must be taken to avoid conversion during processing to a physically metastable, perhaps chemically unstable, form.
2. Metastable crystalline polymorphs may be less chemically stable than the most physically stable crystalline form. Likewise, amorphous drug forms will generally be less chemically stable than the most physically stable form. It is often possible to improve chemical stability of such forms through judicious choice of excipients and formulation processes.
3. If a developer is precluded from developing the lowest energy drug form, for medical benefit or otherwise, it is preferable to develop a stabilized amorphous form, e.g. as a dispersion. Development of a metastable crystalline or amorphous form as a standard physical mixture or granulation with excipients is less preferable, because it is difficult to guarantee that such a formulation will

resist form changes on storage. If the metastable form converts to the stable less soluble form in the dosage form on storage, then *in vivo* C_{max} will almost certainly decrease, and *in vivo* AUC may also decrease depending upon where the drug lies in dose–solubility–permeability space. However, there will be occasional exceptions in which an unstabilized amorphous or metastable crystalline polymorph will be physically stable over the shelf-life of a formulation.

In the end, the manufacturer, whether innovator or generic, must guarantee the quality and bioavailability of the dosage form. It is highly desirable that the drug physical form not change over the storage life of the drug product. If the physical form does change, or if it could change, then the manufacturer must provide assurance (a) that the largest possible change would have no substantive effect on product quality or bioavailability, and/or (b) that extensive scientific study of the formulation guarantees that a change will not occur under all reasonable real-world storage conditions, and/or (c) that analytical methodology and sampling procedures are in place which guarantee that a problem will be detected before dosage forms which have compromised quality or bioavailability can reach patients.

Acknowledgements

We thank Ravi Shanker and Bruno Hancock of Pfizer for very helpful reviews of our manuscript. We also thank Andre Raw of FDA and the anonymous reviewers for comments which helped us communicate our practical perspective with more clarity.

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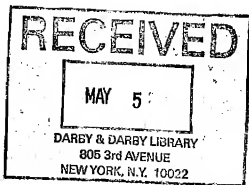
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ISBN: 1-56363-471-6

not due to another thought to induce help predict whether in the future. Although HCG administration the response is tested in children born lotropic hypogonadism (hypothalamic-pituitary deficiency) in pregnancy in the on the onset of the primary ovarian failure pretreated with

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REMERNOR®
(mirtazapine) Tablets

HOW SUPPLIED

REMERNOR® (mirtazapine) Tablets are supplied as:
45 mg Tablets—oval, rounded, yellow, coated, with
"Organ" debossed on one side and "77" on the other side.
Bottles of 30 NDC 0652-0105-30
Bottles of 100 NDC 0652-0105-01
Unit Dose, Box of 100 NDC 0652-0105-00
30 mg Tablets—oval, scored, red-brown, coated, with
"Organ" debossed on one side and "78" on the other side.
Bottles of 30 NDC 0652-0107-30
Bottles of 100 NDC 0652-0107-01
Unit Dose, Box of 100 NDC 0652-0107-00
45 mg Tablets oval, white, coated, with "Organ" de-
bossed on one side and "77" on the other side.
Bottles of 30 NDC 0652-0105-30
Unit dose packs are provided as a blisterpack with 10
tablets each of which contains 10 tablets.

Storage
Store at 25°C (77°F), excursions permitted to 15-30°C
(59-85°F) in USP Controlled Room Temperature. Protect
from light and moisture.

Keep only

Organ,
Manufactured for Organ Inc., West Organ, N.J. 07062
by N.V. Organ, Oss, The Netherlands

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Shown in Product Identification Guide, page 329

REMERNOR SoTab®
(mirtazapine) Tablets

Once-A-Day

DESCRIPTION

REMERNOR SoTab® (mirtazapine) Orally Disintegrating
Tablets are an orally administered drug. Mirtazapine has a
tertiary amine chemical structure and belongs to the piperazine-
azepine group of compounds. It is designated
12,3,4,5-tetrahydro-2-methylpyrido [2,3-b] pyridine
(2,3-b) benzazepine and has the empirical formula of
C₁₆H₁₈N₂. Its molecular weight is 266.36. The structural
formula is the following and it is the racemic mixture:

Mirtazapine is a white to creamy white crystalline powder
which is slightly soluble in water.

REMERNOR SoTab is available for oral administration as
an orally disintegrating tablet containing 15, 30 or 45 mg of
mirtazapine. It disintegrates in the mouth within seconds
after placement on the tongue allowing its contents to be
swallowed with or without water.

REMERNOR SoTab also contains the following inactive in-
gredients: aspartame, citric acid, croscarmellose, hydroxypro-
pyl methylcellulose, magnesium stearate, mannitol, micro-
crystalline cellulose, neoprene and artificial orange flavor,
ethyl-methacrylate, povidone, sodium benzoate, starch, and
talc.

CLINICAL PHARMACOLOGY

Therapeutic Indications

The mechanism of action of REMERNOR SoTab®
(mirtazapine) Orally Disintegrating Tablets, as with other
tricyclic antidepressants in the treatment of major depressive dis-
order, is unknown.

The evidence gathered in preclinical studies suggests that
mirtazapine has central noradrenergic and serotonergic
activity. These studies have shown that mirtazapine acts as
an antagonist at central presynaptic α_1 adrenergic inhibi-
tory autoreceptors and heteroreceptors, an action that is
postulated to result in an increase in central noradrenergic
and serotonergic activity.

Mirtazapine is a potent antagonist of 5-HT₁ and 5-HT₂ re-
ceptors, but has no significant affinity for the 5-HT₁,
5-HT₂, and 5-HT₃ receptors.

Mirtazapine is a potent antagonist of histamine (H₁) recep-
tors, a property that may explain its prominent sedative
effects.

Mirtazapine is a moderate peripheral α_1 adrenergic antag-
onist, a property that may explain the occasional orthostatic
hypotension associated with its use.

Mirtazapine is a moderate antagonist at muscarinic recep-
tors.

Pharmacokinetics
REMERON®/Tab® (mirtazapine) Orally Disintegrating Tablets are rapidly and completely absorbed following oral administration. After a single half-life of about 30–40 hours, peak plasma concentrations are reached. The elimination half-life is about 33 hours in fasting subjects. The presence of food in the stomach has a minimal effect on both the rate and extent of absorption. The elimination half-life is not significantly affected. **REMERON®/Tab®** Orally Disintegrating Tablets are bioequivalent to **REMERON®** (mirtazapine) Tablets.

Metabolism The major metabolic pathway after oral administration and hydrolysis followed by glucuronide conjugation. Major pathways of biotransformation are demethylation and hydrolysis followed by glucuronide conjugation. The major metabolite is the 8-hydroxy metabolite of mirtazapine, which is excreted predominantly via urine (76%) with 15% in feces. Several other metabolites are present in plasma. Mirtazapine is 2 times as long as the (m) enantiomer and therefore achieves a higher AUC than about three times as high as the (s) enantiomer.

Plasma level are linearly related to dose over a dose range of 15 mg to 45 mg. The elimination half-life of mirtazapine is approximately 33 hours in fasting subjects. The elimination half-life after oral administration ranges from 30 to 40 hours across age and gender subgroups, with females of all ages showing slightly longer elimination half-lives than males of all ages. The elimination half-life in elderly males is approximately 37 hours for females vs. 26 hours for males. Steady state plasma levels of mirtazapine are attained within 5 days, with about 50% accumulation.

Excretion Mirtazapine is approximately 85% bound to plasma protein over a concentration range of 0.01–10 µg/mL.

Special Populations

Geriatric
Following oral administration of **REMERON®** (mirtazapine) Tablets 30 mg/day for 7 days to subjects of age 65 and older, the elimination half-life of mirtazapine was reduced in the elderly compared to the younger subjects. The difference was most striking in males, with males, while the clearance of elderly males compared to younger males was reduced by 20–30%. Elderly males were lower compared to younger females. Caution is indicated in administering **REMERON®/Tab®** (mirtazapine) Orally Disintegrating Tablets to elderly patients (see **PRECAUTIONS AND DOSAGE AND ADMINISTRATION**).

Pediatric
The efficacy and effectiveness of mirtazapine in the pediatric population have not been established.

Gender
The mean elimination half-life of mirtazapine after oral administration of the pharmacokinetic study of 30 mg/day across age and gender subgroups, with females of all ages showing slightly longer elimination half-lives than males of all ages (see Table 1). The elimination half-life for females vs. 26 hours for males.

There have been no clinical studies to evaluate the effect of gender on the pharmacokinetics of REMERON®/Tab®.

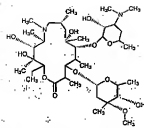
Renal Insufficiency
The disposition of mirtazapine was studied in patients with varying degrees of renal function. Elimination of mirtazapine was reduced in patients with renal impairment. The clearance of mirtazapine was reduced approximately 50% in patients with moderate (Cr = 11–33 mL/min/1.73 m²) and severe (Cr = 3–10 mL/min/1.73 m²) renal impairment when compared to normal subjects. Caution is indicated in administering **REMERON®/Tab®** (mirtazapine) Orally Disintegrating Tablets to patients with compromised renal function (see **PRECAUTIONS AND DOSAGE AND ADMINISTRATION**).

Hepatic Insufficiency
The effect of hepatic impairment on oral dose of **REMERON®** (mirtazapine) Tablets was studied. The clearance of mirtazapine was decreased by approximately 56% in hepatically impaired patients compared to subjects with normal hepatic function. Caution is indicated in administering **REMERON®/Tab®** (mirtazapine) Orally Disintegrating Tablets to patients with compromised hepatic function (see **PRECAUTIONS AND DOSAGE AND ADMINISTRATION**).

Clinical Trials

Efficacy
The efficacy of **REMERON®** (mirtazapine) Tablets as a treatment for major depressive disorder was established in a randomized, 6-week trial in adult outpatients receiving DSM-IV criteria for major depressive disorder. Patients were titrated with mirtazapine from a dose range of 15 mg to 45 mg. Overall, these studies demonstrated that mirtazapine was superior to placebo in the following four measures: 21-Item Hamilton Depression Rating Scale (HDRG) total score; HDRG Depressed Mood Item; HDRG Sleep Item score; and HDRG Appetite and Weight-Related Scale Score (MADRS). Superiority of mirtazapine was also noted for certain features of the HDRG, including the somatic symptom factor and sleep disturbance factor. The mean mirtazapine dose was 30 mg/day. The studies ranged from 21–32 mg/day to 45 mg/day of similar design utilized a higher dose (up to 60 mg/day).

Consult 2004 PDR[®] supplements and future editions for revisions



C₁₆H₁₉N₅O₆
 Serum AUC₀₋₂₄ (μg·hr/mL)
 Serum T_{1/2}

17.4 (2.2)
 71.8 hr

14.9 (3.1)
 68.9 hr

*Total AUC for the entire 5-day and 6-day regimens

AZITHROMYCIN CONCENTRATIONS FOLLOWING
 A 500 mg DOSE (TWO 250 mg CAPSULES) IN ADULTS*

TISSUE OR FLUID	TIME AFTER DOSE (h)	TISSUE OR FLUID CONCENTRATION (μg/g or μg/mL)	CORRESPONDING PLASMA OR SERUM LEVEL (μg/mL)	TISSUE TO PLASMA RATIO
SKIN	72-96	0.4	0.012	35
LUNG	72-96	4.0	0.012	>100
SPUTUM*	2-4	2.4	0.6	2
SPUTUM**	10-12	1.0	0.1	10
TONSIL**	9-18	4.5	0.03	>100
TONSIL**	180	0.9	0.006	>100
CERVIX**	19	2.8	0.04	70

*Azithromycin tissue concentrations were originally determined using 250 mg capsules.

* Sample was obtained 2-4 hours after the first dose.

** Sample was obtained 10-12 hours after the first dose.

*** Dosing regimen of two doses of 250 mg each, separated by 12 hours.

**** Sample was obtained 12 hours after a single 500 mg dose.

starch, sodium croscarmellose, magnesium stearate, sodium lauryl sulfate, hydroxypropyl methylcellulose, lecithin, titanium dioxide, triacetin and D&G Red #30 aluminum lake. ZITHROMAX for oral suspension is supplied in bottles containing azithromycin dihydrate powder equivalent to 300 mg, 500 mg, 900 mg, or 1500 mg azithromycin per bottle and the following inactive ingredients: sucrose, sodium phosphate, triethylamine, hydroxypropyl cellulose, xanthan gum, FD&C Red #40, and spray dried crystallized cherry, cream de vanille and banana flavors. After constitution, each 5 mL of suspension contains 100 mg of 250 mg of azithromycin.

CLINICAL PHARMACOLOGY

Pharmacokinetics
 Following oral administration of a single 500 mg dose (two 250 mg tablets) to 18 fasted healthy male volunteers, the mean (SD) pharmacokinetic parameters were AUC₀₋₂₄ = 4.3 (1.2) μg·hr/mL, C_{max} = 0.5 (0.2) μg/mL, T_{max} = 2.2 (0.9) hours. With a regimen of 500 mg (two 250 mg capsules) on day 1, followed by 250 mg daily (one 250 mg capsule) on days 2 through 5, the pharmacokinetic parameters of azithromycin in plasma in healthy young adults (18-40 years of age) are portrayed in the chart below. C_{max} and C_{min} remained essentially unchanged from day 2 through day 5 of therapy.

Pharmacokinetic Parameters	Total n=12	Day 1	Day 5
(Mean)			
C _{max} (μg/mL)	0.41	0.24	
T _{max} (h)	2.5	1.2	
AUC ₀₋₂₄ (μg·hr/mL)	2.6	2.1	
C _{min} (μg/mL)	0.05	0.05	
Urinary Excret. (% dose)	4.5	6.5	

*Azithromycin 250 mg tablets are bioequivalent to 250 mg capsules in the United States. Azithromycin 250 mg capsules are not commercially available.

In a two-way crossover study, 12 adult healthy volunteers (6 males, 6 females) received 1,500 mg of azithromycin administered in single daily doses for either 5 days (two 250 mg tablets on day 1, followed by one 250 mg tablet on days 2-5) or 3 days (500 mg per day for days 1-3). Due to limited serum samples on day 4 (2-day regimen) and days 2-4 (5-day regimen), the serum concentration-time profile of each subject was fit to a 3-compartment model and the AUC₀₋₂₄ for the fitted concentration profile was comparable between the 5-day and 3-day regimens.

(See first table above)
 Median azithromycin exposure (AUC₀₋₂₄) in mononuclear (MN) and polymorphonuclear (PMN) leukocytes following either the 5-day or 3-day regimen was more than 1,000-fold and 800-fold greater than in serum, respectively. Administration of the same total dose with either the 5-day or 3-day regimen may be expected to provide comparable concentrations of azithromycin within MN and PMN leukocytes.

Two azithromycin 250 mg tablets are bioequivalent to a single 500 mg tablet.

Absorption
 The absolute bioavailability of azithromycin 250 mg capsules is 38%.

In a two-way crossover study in which 12 healthy subjects received a single 500 mg dose of azithromycin (two 250 mg tablets) with or without a high fat meal, food was shown to increase C_{max} by 38% but had no effect on AUC. When azithromycin suspension was administered with food to 28 adult healthy male subjects, C_{max} increased by 56% and AUC was unchanged.

The AUC of azithromycin was unaffected by co-administration of an antacid containing aluminum and magnesium with dextrose with azithromycin capsules, however, the C_{max} was reduced by 24%. Administration of cimetidine (800 mg) two hours prior to azithromycin had no effect on azithromycin absorption.

Distribution
 The serum protein binding of azithromycin is variable in the concentration range approximating human exposure, decreasing from 51% at 0.02 μg/mL to 7% at 2 μg/mL. Following oral administration, azithromycin is widely distributed throughout the body with an apparent steady-state volume of distribution of 31.1 L/kg. Greater azithromycin concentrations in tissues than in plasma or serum were observed. High tissue concentrations should not be inter-

preted to be quantitatively related to clinical efficacy. The antimicrobial activity of azithromycin is pH related and appears to be reduced with decreasing pH. However, the extensive distribution of drug to tissues may be relevant to clinical activity.

Selected tissue (or fluid) concentration and tissue (or fluid) to plasma concentration ratios are shown in the following table.

(See second table above)
 The extensive tissue distribution was confirmed by examination of additional tissues and fluids (nose, sputum, prostate, ovary, uterus, salivary, stomach, liver, and gallbladder). As there are no data from adequate and well-controlled studies of azithromycin treatment of infections in these additional body sites, the clinical importance of these tissue distribution data is unknown.

Following a regimen of 500 mg on the first day and 250 mg daily for 4 days, only very low concentrations were noted in cerebrospinal fluid (less than 0.01 μg/mL) in the presence of non-inflamed meninges.

Metabolism
In vitro and *in vivo* studies to assess the metabolism of azithromycin have not been performed.

Plasma concentrations of azithromycin following single 500 mg oral and *iv* doses declined in a polyphasic pattern with a mean apparent plasma clearance of 550 mL/min and terminal elimination half-life of 58 hours. The prolonged terminal half-life is thought to be due to extensive uptake and subsequent release of drug from tissues.

Biliary excretion of azithromycin, predominantly as unchanged drug, is a major route of elimination. Over the course of a week, approximately 58% of the administered dose appears as unchanged drug in urine.

Special Populations

Renal Insufficiency

Azithromycin pharmacokinetics were investigated in 42 adults (21 to 85 years of age) with varying degrees of renal impairment. Following the oral administration of a single 1,000 mg dose of azithromycin, mean C_{max} and AUC₀₋₂₄ increased by 1.9% and 4.2%, respectively in subjects with mild to moderate renal impairment (GFR 10 to 60 mL/min) compared to subjects with normal renal function (GFR >80 mL/min). The mean C_{min} and AUC₀₋₂₄ increased 61% and 35%, respectively in subjects with severe renal impairment (GFR <10 mL/min) compared to subjects with normal renal function (GFR >80 mL/min). (See DOSAGE AND ADMINISTRATION.)

Hepatic Insufficiency

The pharmacokinetics of azithromycin in subjects with hepatic impairment have not been established.

Gender

There are no significant differences in the disposition of azithromycin between male and female subjects. No dosage adjustment is recommended based on gender.

Geriatric Patients

When studied in healthy elderly subjects aged 60 to 85 years, the pharmacokinetic parameters of azithromycin in elderly men were similar to those in young adults; however, in elderly women, although higher peak concentrations (increased by 30 to 50%) were observed, no significant accumulation occurred.

Pediatric Patients

In two clinical studies, azithromycin for oral suspension was dosed at 10 mg/kg on day 1, followed by 5 mg/kg on days 2 through 6 in two groups of children (ages 1-5 years and 6-15 years, respectively). The mean pharmacokinetic parameters

on day 5 were C_{max} = 0.215 μg/mL (n = 10), T_{max} = 1.0 hour, AUC₀₋₂₄ = 1.822 μg·hr/mL (for the 1- to 5-year-old group) were C_{max} = 0.385 μg/mL, T_{max} = 2.4 hours, AUC₀₋₂₄ = 1.60 μg·hr/mL for the 6- to 15-year-old group. Two clinical studies were conducted in 68 children (ages 6-12 years) to determine the pharmacokinetics and safety of azithromycin for oral suspension in children. Azithromycin was administered following a fast-day breakfast.

The first study consisted of 35 pediatric patients treated with 250 mg tablets (maximum daily dose 500 mg for 8 of whom 14 patients were co-treated for pharmacokinetics). In the second study, 33 pediatric patients received doses of 12 mg/kg/day (maximum daily dose 500 mg for 12 patients). In both studies, azithromycin concentrations were determined over a 24-hour period following the last daily dose. Patients weighing above 35.0 kg in the 8-day study (41.7 kg in the 5-day study) received the same daily dose of 500 mg. Eleven patients (weighing 25.0 kg or less) in the first study and 17 patients (weighing 41.7 kg or less) in the second study received a total dose of 60 mg. The following table shows pharmacokinetic data in the set of children who received a total dose of 60 mg.

Pharmacokinetic Parameter	3-Day Regimen (20 mg/kg × 3 days)	5-Day Regimen (12 mg/kg × 5 days)
n	11	17
C _{max} (μg/mL)	1.1 (0.4)	0.5 (0.4)
T _{max} (hr)	2.7 (1.9)	2.0 (0.8)
AUC ₀₋₂₄ (μg·hr/mL)	7.9 (2.9)	2.9 (1.9)

The similarity of the overall exposure (AUC₀₋₂₄) between 3-day and 5-day regimens in pediatric patients is noteworthy. Single dose pharmacokinetics in children given doses of 30 mg/kg have not been studied. (See DOSAGE AND ADMINISTRATION.)

Drug-Drug Interactions

Drug interaction studies were performed with azithromycin and drugs likely to be co-administered. The effect of co-administration of azithromycin on the pharmacokinetics of other drugs are shown in Table 1 and the effect of other drugs on the pharmacokinetics of azithromycin are shown in Table 2.

Co-administration of azithromycin at therapeutic doses had a modest effect on the pharmacokinetics of the drugs listed in Table 1. No dosage adjustments of drugs listed in Table 1 are recommended when co-administered with azithromycin. Co-administration of azithromycin with theophylline, azithromycin is recommended when administered with azithromycin. Nefazodone significantly increased the C_{max} and AUC of azithromycin. No dosage adjustments of azithromycin are recommended when administered with drugs listed in Table 2. (See PRECAUTIONS - Drug Interactions.)

(See table 1 at bottom of next page.)

(See table 2 at bottom of page 2676.)

Microbiology: Azithromycin acts by binding to the 50S ribosomal subunit of susceptible microorganisms and inhibiting protein synthesis. Azithromycin inhibits protein synthesis in *in vitro* studies.

Azithromycin concentrations in phagocytes and fibroblasts demonstrated by *in vitro* inoculation techniques (discussed in the text of monograph) to extracellular space concentrations was >100 after one hour incubation. *In vivo* studies suggest that concentrations in phagocytes may contribute to drug distribution to inflamed tissues.

Information will be superseded by supplements and subsequent editions

Information will be superseded by supplements and subsequent editions

The safety and efficacy of Weichol® in patients with dyspepsia, swallowing disorders, severe gastrointestinal motility disorders, or major gastrointestinal tract surgery have not been established. Consequently, caution should be exercised when Weichol® is used in patients with these gastrointestinal disorders.

Information for the Patient
Weichol® may be taken once per day with a meal, or taken twice per day in divided doses with meals. Patients should be directed to take Weichol® with a liquid and a meal, and advise them their NCEP-recommended diet. Patients should tell their physicians if they are pregnant, are intending to become pregnant, or are breastfeeding.

Laboratory Tests
Serum total-C, LDL-C, and TG levels should be determined periodically based on NCEP guidelines to confirm favorable initial and adequate long-term responses.

Drug Interactions

Weichol® has been studied in several human drug interaction studies in which it was administered with a meal and the test drug. Weichol® was found to have no significant effect on the bioavailability of digoxin, losartan, metoprolol, quindine, valproic acid, and warfarin. Weichol® decreased the C_{max} and AUC of sustained-release verapamil (Calan SR) by approximately 31% and 11%, respectively. Since there is a high degree of variability in the bioavailability of verapamil, the clinical significance of this finding is unclear. In clinical studies, co-administration of Weichol® with atorvastatin, lovastatin, or simvastatin did not interfere with the lipid-lowering activity of the HMG-CoA reductase inhibitors. Other drugs have not been studied. When administering other drugs for which alterations in blood levels could have a clinically significant effect on safety or efficacy, physicians should consider monitoring drug levels or effects.

Contraception, Mutagenesis, Impairment of Fertility
A 104-week carcinogenicity study with celecoxib (Weichol®) was conducted in CD-1 mice, at oral dosages of 50 mg/kg/day. This dose was approximately 50 times the maximum recommended human dose of 4.5 g/day based on body weight, mg/kg. There were no significant drug-induced tumor findings in male or female mice. In a 104-week carcinogenicity study with celecoxib (Weichol®) in Harlan Sprague-Dawley rats, a statistically significant increase in the incidence of pancreatic acinar cell adenoma was seen in male rats at doses ≥ 1.5 g/kg/day (approximately 50 times the maximum human dose, based on body weight, mg/kg) (trend test only). A statistically significant increase in thyroid C-cell adenoma was seen in female rats at 2.4 g/kg/day (approximately 40 times the maximum human dose, based on body weight, mg/kg). Celecoxib and four degradants present in the drug substance have been evaluated for mutagenicity in the Ames test and a mammalian chromosomal aberration test. The four degradants and an extract of the parent compound did not exhibit genetic toxicity in an *in vitro* bacterial mutagenesis assay in *S. typhimurium* and *E. coli* (Ames assay) with or without rat liver metabolic activation. An extract of the parent compound was positive in the Chinese Hamster Ovary (CHO) cell chromosomal aberration assay in the presence of metabolic activation and negative in the absence of metabolic activation. The results of the CHO cell chromosomal aberration assay with two of the four degradants, dextroisomer HCl and enantiomerically pure ammonium chloride HCl, were equivocal in the absence of metabolic activation and negative in the presence of metabolic activation. The other two degradants, dextroisomer HCl and 6-dehydroisomerically pure ammonium chloride HCl, were negative in the presence and absence of metabolic activation.

Celecoxib did not impair fertility in rats at doses of up to 3 g/kg/day (approximately 50 times the maximum human dose, based on body weight, mg/kg).

PREGNANCY
Pregnancy Category B
Reproduction studies have been performed in rats and rabbits at doses up to 3 g/kg/day and 1 g/kg/day, respectively (approximately 50 and 17 times the maximum human dose, based on body weight, mg/kg) and have revealed no evidence of harm to the fetus or to celecoxib. There are, however, no adequate and well-controlled studies in pregnant women. Because animal reproduction studies are not always predictive of human response, this drug should be used during pregnancy only if clearly needed. Requirements for vitamins and other nutrients are increased in pregnancy. The effect of Weichol® on the absorption of vitamins has not been studied in pregnant women.

Pediatric Use

The safety and efficacy of celecoxib (Weichol®) have not been established in pediatric patients.

Geriatric Use

There is no evidence for special considerations when celecoxib (Weichol®) is administered to elderly patients.

ADVERSE REACTIONS

Weichol® treatment-emergent adverse events that occurred in greater than 2% of patients in an integrated safety analysis are presented in Table 4.

RISK CATEGORY	LDL-C GOAL	TO INCREASE THERAPEUTIC LIFESTYLE CHANGES (TLC)	TO CONSIDER DRUG THERAPY
CHD or CHD Risk Equivalents (10-year risk >20%)	<100 mg/dL	≥ 100 mg/dL	≥ 180 mg/dL (100-180 mg/dL drug optional) ^a
2+ Risk Factors (10-year risk $\geq 20\%$)	<130 mg/dL	≥ 130 mg/dL	10-year risk 10-20% ≥ 130 mg/dL
0-1 Risk Factor	<160 mg/dL	≥ 160 mg/dL	10-year risk <10% ≥ 160 mg/dL
			100-180 mg/dL LDL lowering drug optional ^a

^aSome authorities recommend use of LDL cholesterol-lowering drugs in the category if LDL cholesterol < 100 mg/dL cannot be achieved by therapeutic lifestyle changes. Other products of drugs that primarily affect triglycerides and/or cholesterol ≥ 0.5 , niacin acid or fibrate. Clinical judgment also may call for deferring drug therapy in this category. ^bAlmost all people with 0-1 risk factor have a 10-year risk $< 10\%$, thus 10-year risk assessment in people with 0-1 risk factor is not necessary.

Table 4: Frequent (>2%) Treatment-Emergent Adverse Events by Treatment Category

Body as a Whole	ADVERSE EVENT	PLACED (N=258) %	WEICHOL® ONLY (N=907) %
Infection	18	10	
Headache	8	6	
Pain	7	5	
Back Pain	6	3	
Abdominal Pain	5	5	
Flu Syndrome	3	3	
Accidental Injury	3	4	
Anthrax	2	4	
Digestive System			
Flatulence	14	12	
Constipation	7	11	
Diarrhea	7	5	
Nausea	4	4	
Dyspepsia	3	8	
Respiratory System			
Sinusitis	4	2	
Rhinitis	3	3	
Cough Increased	2	2	
Pharyngitis	2	3	
Musculoskeletal System			
Myalgia	0	2	

OVERDOSAGE

Because Weichol® is not absorbed, the risk of systemic toxicity is low. Doses in excess of 4.5 g per day have not been tested.

DOSE AND ADMINISTRATION

Monotherapy

The recommended starting dose of Weichol® is 3 tablets taken twice per day with meals or 6 tablets once per day with a meal. The Weichol® dose can be increased to 7 tablets, depending upon the desired therapeutic effect. Weichol® should be taken with a liquid.

Combination Therapy

Weichol®, at doses of 4 to 6 tablets per day, has been shown to be safe and effective when dosed at the same time (i.e., co-administered) as an HMG-CoA reductase inhibitor or when two drugs are dosed apart. (CLINICAL PHARMACOLOGY, Clinical Effects). Weichol® should be taken with a liquid. For maximal therapeutic effect in combination with an HMG-CoA reductase inhibitor, the recommended dose of Weichol® is 3 tablets taken twice per day with meals or 6 tablets taken once per day with a meal.

HOW SUPPLIED

Weichol® (celecoxib hydrochloride), 625 mg, is supplied as an off-white, solid tablet imprinted with the word "Sanofi" over "COI".

Weichol® tablets are available as follows:

Bottles of 180—NDC 65597-701-18

Bottles of 24—NDC 65597-701-24

Storage

Store at 20°C (77°F); excursions permitted to 15-30°C (59-86°F) [see USP Controlled Room Temperature]. Do not expose to 40°C or do not adversely affect the product. Protect from moisture.

References

- Grundy SM, Albers CE, Selen G. Interruption of the therapeutic circulation of bile acids in man: comparative effects of cholestyramine and diet exclusion on cholesterol metabolism. *J Lab Clin Med* 1971; 78: 94-122.
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- Friedewald WT, Levy RI, Fredrickson DS. Estimating the concentration of LDL cholesterol in plasma without use of a preparative ultracentrifuge. *Clin Chem* 1972; 18: 499.

Manufactured by: Sanofi Pharm Inc.

Parishan, New Jersey 07054

by: Pathon YM Inc. MSB 175

Thomson, Ontario, Canada M5B 1Y5

Licensed From: Glaxo Pharmaceuticals, Inc.

Issued: March 2005

Version: 6

Shown in Product Identification Guide, page 354

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ARXITRA®

(in rox®-erol)

(fendapirone sodium) injection

For full prescribing information, please see Organo Sanofi-Synthelabo LLC.

AMBIBEN®

(am-bi-ben)

(tolpimide tartrate)

DESCRIPTION

Ambien (zolpidem tartrate), is a non-benzodiazepine hypnotic of the imidazopyridine class and is available in 5-mg and 10-mg strength tablets for oral administration. Chemical structure is N,N'-dimethyl-2-pyridine-3-ylmethyl-2,3-pyridine-3-carboxamide 1,1'-di-*tert*-butyl ester (2:1). It has the following structure:

See chemical structure at top of column.

Zolpidem tartrate is a white to off-white crystalline powder that is sparingly soluble in water, ethanol, and propylene glycol. It has a molecular weight of 764.84.

Each Ambien tablet includes the following inactive ingredients: hydroxypropyl methylcellulose, lactose, magnesium stearate, microcrystalline cellulose, polyethylene glycol,

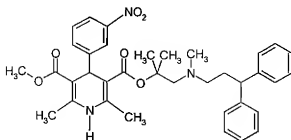
EXHIBIT D

ZANIDIP PRODUCT INFORMATION (lercanidipine tablets)

DESCRIPTION

Lercanidipine hydrochloride.

Lercanidipine is a dihydropyridine derivative. It is a racemate due to the presence of a chiral carbon atom at position 4 of the 1,4-dihydropyridine ring.



Chemical name: 3,5-pyridinedicarboxylic acid, 1,4- dihydro-2, 6-dimethyl-4-(3-nitrophenyl)-2-[[[3,3-diphenylpropyl)methylamino]-1,1-dimethylethyl methyl ester hydrochloride. MW: 648.2 (free base: 611.7).

Lercanidipine hydrochloride (CAS: 132866-11-6) is a microcrystalline, odourless, citrine powder, readily soluble in chloroform and methanol, practically insoluble in water. Octanol:water partition coefficient (LogP): 6.4.

Zanidip tablets also contain the excipients lactose, microcrystalline cellulose, sodium starch glycollate, povidone and magnesium stearate. The tablets are film-coated with the proprietary ingredients Opadry OY-SR-6497 (10 mg-yellow) or Opadry O2-F2-5077 (20 mg-pink).

PHARMACOLOGY

Pharmacodynamic Properties

Lercanidipine is a calcium antagonist of the dihydropyridine group and selectively inhibits the transmembrane influx of calcium into cardiac and vascular smooth muscle, with a greater effect on vascular smooth muscle than on cardiac smooth muscle. The antihypertensive action is due to a direct relaxant effect on vascular smooth muscle which lowers total peripheral resistance and hence blood pressure. Lercanidipine has a prolonged antihypertensive activity because of its high membrane partition coefficient. It is devoid of negative inotropic effects and its vascular selectivity is due to its voltage-dependent calcium antagonist activity. Since the vasodilatation induced by lercanidipine hydrochloride is

gradual in onset, acute hypotension with reflex tachycardia has rarely been observed in hypertensive patients. As for other asymmetric 1,4-dihydropyridines, the antihypertensive activity of lercanidipine is mainly due to the (S) – enantiomer. No significant effects on ECG have been seen.

Clinical Trials

Placebo-controlled studies

Lercanidipine has been compared to placebo in four (4) to 16-week studies, involving 671 patients with mild-moderate essential hypertension. All studies used a 3-week placebo run-in period. Endpoints were diastolic and systolic blood pressure 24 hours post dose. Both 10mg and 20mg once daily significantly lowered diastolic and systolic blood pressure compared to placebo, and the reduction in blood pressure was maintained throughout the 24 hour dosing period. Diastolic blood pressure changes observed after 4-week treatment with 10-20 mg QD lercanidipine ranged between 8 and 13 mmHg, as compared to 3-6 mmHg induced by placebo. Studies with 24-hour ambulatory blood pressure monitoring have documented that the antihypertensive effect of lercanidipine is maintained throughout the 24 hour dosing period, with limited variations between peak (5-7 hours post dosing) and trough blood pressure changes.

Active-controlled studies

Four clinical trials involving 538 patients with mild-moderate essential hypertension have compared lercanidipine with nifedipine SR, atenolol, hydrochlorothiazide and captopril. Trials included a 2-week washout period followed by a 3-week placebo run-in, and 12-16 weeks of active treatment. Diastolic and systolic blood pressure was measured 24 hours post-dose. Lercanidipine was as effective as the comparator drugs, and at least as well tolerated. 24-hour blood-pressure monitoring was used in a comparative, cross-over trial of lercanidipine versus amlodipine (n=16). The effect of lercanidipine paralleled that of amlodipine throughout the 24 hour period.

Patients with Isolated Systolic Hypertension

The effect of lercanidipine 10-20mg daily on isolated systolic hypertension was studied in a double-blind, randomised, placebo-controlled study in 83 elderly patients (sitting SBP>160mm Hg and sitting DBP<95mm Hg). The study consisted of 1 week wash-out, 3 weeks placebo run-in, and 8 weeks of active treatment. Systolic and diastolic blood pressure was measured 24 hours post dose. Lercanidipine 10 to 20 mg was efficacious in lowering systolic blood pressure from the initial values of 172.6± 5.6 mmHg to 140.2± 8.7mmHg (mean±SD, per-protocol population in all patients completing the whole 8 weeks of double-blind treatment), as compared to the changes in the placebo group (from 172.4±6.3 to 162.8±9.7 mmHg). Therefore, at study endpoint, patients treated with lercanidipine experienced a significantly greater decrease (-22.6mm Hg, p<0.001) in sitting systolic blood pressure in comparison to placebo. The diastolic blood pressure was within normal range.

Long-term studies

In long term studies, 399 patients were followed for 12 months, with dose titration allowed every 4 weeks, to 30mg daily. Development of tolerance was not seen. The antihypertensive effect was maintained, and the heart rate was not significantly affected. A further fall in blood pressure was seen after the first and second month, with blood pressure stabilising in the third month. The majority of patients remained on 10mg once daily.

Pharmacokinetics**Absorption**

Lercanidipine is completely absorbed after oral administration. Peak plasma levels of $3.30\text{ng/mL} \pm 2.09\text{ s.d}$ and $7.66\text{ ng/mL} \pm 5.90\text{ s.d}$ occur 1.5-3 hours after dosing with 10mg and 20mg, respectively. The absolute bioavailability of lercanidipine is about 10%, because of high first pass metabolism. The bioavailability increases 4-fold when lercanidipine is ingested up to 2 hours after a high fat meal, and about 2-fold when taken immediately after a carbohydrate-rich meal. Consequently, lercanidipine should be taken at least 15 minutes before a meal.

With oral administration, lercanidipine exhibits non-linear kinetics. After 10, 20 or 40mg, peak plasma concentrations observed were in the ratio 1:3:8 and areas under plasma concentration-time curves in the ratio 1:4:18, showing a progressive saturation of first pass metabolism.

Accordingly, bioavailability increases as dosage increases.

The two enantiomers of lercanidipine have a similar time to peak plasma concentration. The peak plasma concentration and AUC are, on average, 1.2-fold higher for the (S) enantiomer. No *in vivo* interconversion of enantiomers is observed.

Distribution

Distribution of lercanidipine from plasma to tissues and organs is rapid and extensive. Serum protein binding exceeds 98%. The free fraction of lercanidipine may be increased in patients with renal or hepatic impairment as plasma protein levels are decreased in these disease states.

Metabolism

As for other dihydropyridine derivatives, lercanidipine is extensively metabolised by CYP3A4. It is predominantly converted to inactive metabolites; no parent drug is found in the urine or faeces. About 50% of the dose is excreted in the urine.

Elimination

The mean terminal elimination half-life of S- and R-lercanidipine enantiomers is 5.8 ± 2.5 and 7.7 ± 3.8 hours, respectively. No accumulation was seen upon repeated administration. The therapeutic activity of lercanidipine lasts for 24 hours, due to its high binding to lipid membranes.

Elderly patients

In elderly patients, the pharmacokinetics of lercanidipine is similar to that observed in the general population.

Hepatic Impairment

A study in patients with mild hepatic impairment (Child-Pugh class A) showed that the pharmacokinetic behaviour of the drug is similar to that observed in the general population. No studies have been undertaken in patients with moderate or severe hepatic impairment.

Renal impairment

In patients with severe renal dysfunction (creatinine clearance < 12 mL/min) or dialysis-dependent patients, plasma levels were increased by about 70%. As a consequence, the drug should be contraindicated in severe renal impairment.

INDICATIONS

Zanidip is indicated for the treatment of hypertension.

CONTRAINDICATIONS

- Hypersensitivity to any dihydropyridine or any ingredient of Zanidip;
- Severe hepatic impairment;
- Severe renal impairment (creatinine clearance < 12 mL/min).
- Concomitant treatment of Zanidip with cyclosporin should be avoided

PRECAUTIONS

Ischaemic heart disease

It has been suggested that some short-acting dihydropyridines may be associated with increased cardiovascular risk in patients with ischaemic heart disease. Although lercanidipine is long-acting, caution should be required in such patients.

Outflow obstruction (aortic stenosis)

Lercanidipine should be administered with caution in patients with left ventricular outflow obstruction (aortic stenosis).

Congestive heart failure

In general calcium channel blockers should be used with caution in patients with heart failure. Although animal data and acute haemodynamic evaluation in patients with preserved left ventricular function have not demonstrated that lercanidipine exerts a direct negative inotropic effect, safety in patients with congestive heart failure has not been established. Therefore, as for other calcium channel blockers, lercanidipine should be used with caution in such patients, especially if untreated.

Unstable angina pectoris or within one month of a myocardial infarction

Rarely patients have developed documented increased frequency, duration and/or severity of angina on starting calcium channel blocker therapy or at the time of dosage increase (particularly those with severe obstructive coronary artery disease). The mechanism of this effect has not been elucidated, however the possibility of an exacerbation of angina and/or cardiac ischaemia exists. It is therefore suggested that the use of

calcium channel blockers is not advisable in patients with unstable angina pectoris or recent myocardial infarction.

Carcinogenesis, mutagenesis, impairment of fertility

No evidence for genotoxic activity was observed with lercanidipine in *in vitro* assays of gene mutation (reverse mutation in *S. Typhimurium*, forward mutation in Chinese Hamster V79 fibroblasts), gene conversion (in *saccharomyces cerevisiae* D4) or chromosomal damage (CHO cytogenetic assay). Negative findings were also obtained with lercanidipine in an *in vivo* assay of chromosomal damage (mouse micronucleus test).

Carcinogenicity studies of lercanidipine (administered via the diet) have been performed in rats and mice (18 months), using doses up to 60 mg/kg/day for mice and 5 mg/kg/day for rats. Plasma concentrations (AUC) of lercanidipine at the highest doses used in these studies were about 2-4 times the highest AUC expected in humans during treatment with lercanidipine. Lercanidipine showed no evidence of carcinogenic activity in these studies.

Administration of lercanidipine at oral doses up to 12 mg /kg /day (associated with plasma lercanidipine concentration (AUC) about 20-40 times higher than the expected human AUC) had no effect in male or female fertility in rat.

Use in pregnancy: Category C

There is no clinical experience with lercanidipine in pregnancy, but other dihydropyridine compounds have been found to cause irreversible malformations in animals. Therefore, lercanidipine should not be administered during pregnancy or to women with child-bearing potential unless effective contraception is used.

In animal studies, pregnant rats given lercanidipine orally at doses \geq 2.5 mg/kg/day, beginning prior to mating, or 12 mg/kg/day, beginning from early gestation, showed signs of distocia and had a increased incidence of still births and a lower neonatal survival index. The no-effect dose for effects on parturition and neonatal survival was 0.5 mg/kg/day (associated with lercanidipine concentration (AUC) about 50% of the expected human AUC) when dosing started before pregnancy or 2.5 mg/kg/day (about 3 times the human AUC) when dosing started during early gestation. Administration with lercanidipine at doses of 2.5 mg/kg/day during gestation also caused a higher incidence of fetal visceral abnormalities (mono/bilateral renal pelvic and/or ureteric dilatation) and skeletal abnormalities (mainly delayed ossification) at all dose levels. A no-effect dose was not established. The effects of lercanidipine during pregnancy have not been investigated adequately in a non-rodent species.

Use in lactation

There is no clinical experience with lercanidipine in lactation. Distribution into milk may be expected, due to the high lipophilicity of lercanidipine. Therefore, lercanidipine should not be administered to lactating women.

Use in the elderly

Although the pharmacokinetic data and clinical experience suggest that no adjustment of the daily dose is required, special care should be exercised when initiating treatment in the elderly.

Use in children

Due to lack of clinical experience, lercanidipine is not recommended for use in patients under the age of 18.

Use in hepatic impairment

The pharmacokinetics of lercanidipine in patients with mild hepatic impairment are similar to those observed in the general population. However, there are no studies in patients with moderate hepatic impairment and dosage recommendations have not been established. Lercanidipine should therefore be used with caution in this patient group and careful monitoring undertaken during treatment, since the bioavailability and hypotensive effect may be increased. The use of Lercanidipine in patients with moderate hepatic impairment should only be undertaken if the benefits are considered to outweigh the risks. Lercanidipine is contraindicated, in patients with severe hepatic disease.

Use in renal impairment

Although the pharmacokinetics of lercanidipine in patients with mild to moderate renal impairment are similar to those observed in the general population, special care should be exercised when commencing the treatment in such patients. The usual recommended dose of 10mg daily may be tolerated; however, an increase to 20mg daily should be approached with caution.

Interaction with other drugs

Lercanidipine has been safely administered with diuretics and ACE inhibitors. It may also be administered safely with beta-blockers which are eliminated unchanged (such as atenolol).

Inhibitors or inducers of Cytochrome CYP3A4

Since the main metabolic pathway of lercanidipine involves the enzyme CYP3A4, drugs that inhibit or induce this enzyme have the potential to alter the plasma concentration of the compound.

Therefore, inhibitors of CYP3A4 (such as ketoconazole, itraconazole, erythromycin, ritonavir and fluoxetine) may increase the plasma concentration of lercanidipine, and such combinations should be used with caution.

When co-administered with CYP3A4 inducers, such as anticonvulsants (eg. phenytoin, carbamazepine) and rifampicin, the antihypertensive effect of lercanidipine may be reduced and, therefore, blood pressure should be monitored when the co-administration is foreseen.

CYP3A4 and CYP2D6 substrates

The potential for *in vivo* inhibition of CYP3A4 by lercanidipine is negligible, as confirmed by an interaction study with midazolam in healthy volunteers. After repeated co-administration with lercanidipine, midazolam (a probe for CYP3A4 activity) was found to be essentially bioequivalent to the drug administered alone. However, unless specific data are available, caution should also be exercised when lercanidipine is co-prescribed with other substrates of CYP3A4 which have a narrow therapeutic index, such as cyclosporin, and class III antiarrhythmic drugs (e.g. amiodarone and quinidine).

Co-administration of lercanidipine with cyclosporin resulted in a 3 fold increase in the plasma levels of lercanidipine and a 21% increase in the bioavailability of cyclosporin. However, when cyclosporin was administered 3 hours after lercanidipine, no increase in plasma levels was observed for lercanidipine, while the bioavailability of cyclosporin

increased by 27%. Therefore, cyclosporin and lercanidipine should not be administered together.

Moreover, interaction studies in humans have shown that lercanidipine did not modify the plasma levels of metoprolol, (a typical substrate of CYP2D6). Therefore, at therapeutic doses it is unlikely that lercanidipine will inhibit the biotransformation of drugs metabolized by CYP2D6. These findings confirm that the inhibition of cytochrome P450 isoenzymes observed *in vitro* with lercanidipine is devoid of any clinical significance. *In vitro* experiments with human liver microsomes demonstrated that lercanidipine inhibits CYP3A4 and CYP2D6 (IC₅₀ of 2.6 μ m and 0.8 μ m, respectively). The IC₅₀ concentrations for CYP3A4 and CYP2D6 are 160 and 40 fold higher, respectively, than those reached at peak in the plasma after a 20mg dose.

Beta-blockers

When lercanidipine was administered with metoprolol, a beta-blocker eliminated mainly by the liver, the bioavailability of metoprolol was not changed, while that of lercanidipine was reduced by 50%. Therefore, when co-administered with metoprolol, it may be necessary to increase the dose of lercanidipine. It is anticipated that a similar effect may occur with propranolol.

Cardiac glycosides

Co-administration of lercanidipine in patients chronically treated with beta-methyl digoxin (a pro-drug of digoxin) showed no evidence of a pharmacokinetic interaction. However, patients on concomitant digoxin treatment should be closely monitored.

Cimetidine

Concomitant administration of cimetidine 400mg BD does not cause significant changes in the plasma levels of lercanidipine: AUC and C_{max} were increased by a mean of 11%. However, at higher doses caution is required since the bioavailability and the hypotensive effect of lercanidipine may be increased.

Simvastatin

Co-administration of a 20 mg dose of lercanidipine with 40 mg simvastatin resulted in no increase in the bioavailability of lercanidipine, however a 56% increase was observed for simvastatin and a 28% increase for its active metabolite β -hydroxyacid. It is unlikely that these changes are clinically relevant. However, it is recommended that when required lercanidipine be administered in the morning and simvastatin in the evening.

Food

See previous section on pharmacokinetics.

The metabolism of dihydropyridines can be inhibited by grapefruit juice, leading to increased plasma concentration and hypotensive effect.

Alcohol should be avoided while taking lercanidipine since it may potentiate the effect of vasodilating antihypertensive drugs.

ADVERSE REACTIONS

Treatment with lercanidipine is generally well tolerated. In nine placebo-controlled clinical trials with a treatment duration lasting at least 4 weeks, 582 patients were initially treated with lercanidipine, and 292 patients received placebo. Most of the events reported in the studies were related to the vasodilatory effects of lercanidipine, and were classified mild-moderate in severity.

Table 1 lists, according to organ system, adverse events that were reported in placebo controlled trials in hypertensive patients with lercanidipine tablets at an incidence greater than or equal to 1% in at least one of the active treatment groups.

Table 1

Adverse Event	Lercanidipine 10mg once daily (%)	Lercanidipine 20mg once daily (titrated) (%)	Placebo (%)
CARDIOVASCULAR			
Flushing	2.6	2.2	1.6
Palpitations/Tachycardia	1.5	1.1	0.3
BODY AS A WHOLE			
Peripheral oedema	1.0	1.1	0.9
CENTRAL & PERIPHERAL NERVOUS SYSTEM			
Dizziness	1.0	0.0	0.6
Headache	4.4	4.3	2.5
LIVER DISORDERS			
GGT increased	0.0	1.1	0.3

More extensively, over 15500 patients were treated with lercanidipine in clinical trials (including PMS studies) with doses from 2.5 mg QD up to 40 mg QD, and with treatment duration ranging from single dose up to more than 1 year. Adverse experiences which were not clearly drug related and which occurred in <1% but \geq 0.1% of patients are summarized according to organ system.

Cardiovascular: palpitations/tachycardia.

Central and Peripheral nervous system: dizziness, vertigo.

Gastrointestinal: nausea, dyspepsia, abdominal pain, diarrhoea.

Psychiatric: somnolence.

General: flushing, asthenia (including fatigue and muscle weakness).

The following events have been rarely reported:

Cardiovascular: hypotension, orthostatic hypotension, periorbital oedema, anginal pain, myocardial infarction, cardiac failure.

Respiratory: dyspnoea.

Central and Peripheral nervous system: migraine, paraesthesia, cramps legs.

Special senses: taste alteration.

Gastrointestinal: vomiting, GI disorder NOS.

Liver and biliary system: GGT increased.

Genitourinary: polyuria, urinary frequency, impotence.

Musculoskeletal: myalgia.

Skin and appendages: rash, pruritus, allergic dermatitis, hives, sweating increased.

Psychiatric: anxiety, insomnia.

Metabolic: Hypercholesterolaemia.

General: chest pain, malaise.

Serious adverse events have been reported in clinical trials in less than 0.002% of the patients. The remaining adverse events have been reported as mild to moderate in intensity.

Laboratory tests

There were reports of isolated and reversible increases in serum levels of hepatic transaminases; no other clinically significant pattern of laboratory test abnormalities related to lercanidipine has been observed. Lercanidipine does not effect blood sugar or lipid levels.

DOSAGE AND ADMINISTRATION

The recommended dose is 10mg once daily, at least 15 minutes before a meal. The dose may be increased to 20mg once daily depending on the individual response. Dose titration should be gradual, as it may take about 2 weeks for the maximal antihypertensive effect to be apparent. Titration may proceed more rapidly, however, if clinically warranted, provided the patient is assessed frequently. Since it is unlikely that increasing the dose beyond 20mg will further improve the efficacy, and may be associated with side effects, doses above 20 mg are not recommended. Some individuals not adequately controlled on a single antihypertensive agent may benefit from the addition of lercanidipine at the same doses used in monotherapy to the existing regimen with a beta-blocker, a diuretic or an ACE-inhibitor.

Use in elderly, children, hepatic and renal impairment: see precautions.

OVERDOSAGE

There is no experience with lercanidipine overdosage. As with other dihydropyridines, overdosage might be expected to cause excessive peripheral vasodilation with marked hypotension and reflex tachycardia. In case of severe hypotension, bradycardia and unconsciousness, cardiovascular and respiratory monitoring will be required, and supportive treatment may be necessary. Since lercanidipine is highly lipophilic, dialysis is unlikely to be effective.

PRESENTATION

ZANIDIP is available as 10 mg or 20 mg tablets.

10 mg: Yellow, round, scored, film-coated tablets, containing lercanidipine 9.4 mg (present as 10mg of lercanidipine hydrochloride).

20 mg: Pink, circular, biconvex, film-coated tablets, containing lercanidipine 18.8 mg (present as lercanidipine hydrochloride 20 mg).
Packs of 7 or 30 tablets.

STORAGE

Store below 30 degrees Celsius. Protect from moisture and light.

NAME AND ADDRESS OF THE SPONSOR

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DATE OF TGA APPROVAL

Approved by Therapeutic Goods Administration: 16 December 2005